

Package ‘PLNmodels’

March 16, 2021

Title Poisson Lognormal Models

Version 0.11.4

Description The Poisson-lognormal model and variants (Chiquet, Mariadassou and Robin, 2020 <doi:10.1101/2020.10.07.329383>) can be used for a variety of multivariate problems when count data are at play, including principal component analysis for count data, discriminant analysis, model-based clustering and network inference. Implements variational algorithms to fit such models accompanied with a set of functions for visualization and diagnostic.

URL <https://pln-team.github.io/PLNmodels/>

BugReports <https://github.com/pln-team/PLNmodels/issues>

License GPL (>= 3)

Encoding UTF-8

RoxygenNote 7.1.1

Depends R (>= 3.4)

LazyData true

biocViews

Imports methods, stats, MASS, future, future.apply, R6, glassoFast, Matrix, Rcpp, nloptr (>= 1.2.0), igraph, grid, gridExtra, dplyr, tidyr, purrr, ggplot2, corrplot, magrittr, rlang

Suggests knitr, rmarkdown, testthat, covr, pkgdown, biomformat, phyloseq, spelling, factoextra, Wrench

LinkingTo Rcpp, RcppArmadillo, nloptr (>= 1.2.0)

VignetteBuilder knitr

Collate 'PLNfit-class.R' 'PLN.R' 'PLNLDA.R' 'PLNLDAfit-S3methods.R' 'PLNLDAfit-class.R' 'PLNPCA.R' 'PLNPCAfamily-S3methods.R' 'PLNfamily-class.R' 'PLNPCAfamily-class.R' 'PLNPCAfit-S3methods.R' 'PLNPCAfit-class.R' 'PLNfamily-S3methods.R' 'PLNfit-S3methods.R' 'PLNmixture.R' 'PLNmixturefamily-S3methods.R' 'PLNmixturefamily-class.R' 'PLNmixturefit-S3methods.R' 'PLNmixturefit-class.R'

'PLNmodels-package.R' 'PLNnetwork.R'
 'PLNnetworkfamily-S3methods.R' 'PLNnetworkfamily-class.R'
 'PLNnetworkfit-S3methods.R' 'PLNnetworkfit-class.R'
 'RcppExports.R' 'deprecated.R' 'import_utils.R' 'mollusk.R'
 'oaks.R' 'plot_utils.R' 'trichoptera.R' 'utils-pipe.R'
 'utils.R' 'zzz.R'

Language en-US

NeedsCompilation yes

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Repository CRAN

Date/Publication 2021-03-16 16:10:02 UTC

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coef.PLNfit

*Extract model coefficients***Description**

Extracts model coefficients from objects returned by `PLN()` and its variants

Usage

```
## S3 method for class 'PLNfit'
coef(object, type = c("main", "covariance"), ...)
```

Arguments

`object` an R6 object with class `PLNfit`
`type` type of parameter that should be extracted. Either "main" (default) for

$$\Theta$$

or "covariance" for

$$\Sigma$$

... additional parameters for S3 compatibility. Not used

Value

A matrix of coefficients extracted from the PLNfit model.

See Also

[sigma.PLNfit\(\)](#), [vcov.PLNfit\(\)](#), [standard_error.PLNfit\(\)](#)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1 + offset(log(Offset)), data = trichoptera)
coef(myPLN) ## Theta
coef(myPLN, type = "covariance") ## Sigma
```

coef.PLNLDAfit	<i>Extracts model coefficients from objects returned by PLNLDA()</i>
----------------	--

Description

The method for objects returned by [PLNLDA\(\)](#) only returns coefficients associated to the

$$\Theta$$

part of the model (see the PLNLDA vignette for mathematical details).

Usage

```
## S3 method for class 'PLNLDAfit'
coef(object, ...)
```

Arguments

object	an R6 object with class PLNLDAfit
...	additional parameters for S3 compatibility. Not used

Value

Either NULL or a matrix of coefficients extracted from the PLNLDAfit model.

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLNLDA <- PLNLDA(Abundance ~ Wind, grouping = Group, data = trichoptera)
coef(myPLNLDA)
```

 coef.PLNmixturefit *Extract model coefficients*

Description

Extracts model coefficients from objects returned by [PLN\(\)](#) and its variants

Usage

```
## S3 method for class 'PLNmixturefit'
coef(object, type = c("main", "means", "covariance", "mixture"), ...)
```

Arguments

object	an R6 object with class PLNmixturefit
type	type of parameter that should be extracted. Either "main" (default) for Θ , "means" for μ , "mixture" for π , or "covariance" for Σ
...	additional parameters for S3 compatibility. Not used

Value

A matrix of coefficients extracted from the PLNfit model.

See Also

[sigma.PLNmixturefit\(\)](#)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLNmixture(Abundance ~ 1 + offset(log(Offset)),
  data = trichoptera, control_main = list(iterates = 0)) %>% getBestModel()
coef(myPLN) ## Theta - empty here
coef(myPLN, type = "mixture") ## pi
coef(myPLN, type = "means") ## mu
coef(myPLN, type = "covariance") ## Sigma
```

coefficient_path	<i>Extract the regularization path of a PLNnetwork fit</i>
------------------	--

Description

Extract the regularization path of a PLNnetwork fit

Usage

```
coefficient_path(Robject, precision = TRUE, corr = TRUE)
```

Arguments

Robject	an object with class <code>PLNnetworkfamily</code> , i.e. an output from <code>PLNnetwork()</code>
precision	a logical, should the coefficients of the precision matrix Omega or the covariance matrix Sigma be sent back. Default is TRUE.
corr	a logical, should the correlation (partial in case <code>precision = TRUE</code>) be sent back. Default is TRUE.

Value

Sends back a tibble/data.frame.

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
head(coefficient_path(fits))
```

compute_offset	<i>Compute offsets from a count data using one of several normalization schemes</i>
----------------	---

Description

Computes offsets from the count table using one of several normalization schemes (TSS, CSS, RLE, GMPR, etc) described in the literature.

Usage

```
compute_offset(
  counts,
  offset = c("TSS", "GMPR", "RLE", "CSS", "Wrench", "none"),
  ...
)
```

Arguments

counts	Required. An abundance count table, preferably with dimensions names and species as columns.
offset	Optional. Normalization scheme used to compute scaling factors used as offset during PLN inference. Available schemes are "TSS" (Total Sum Scaling, default), "CSS" (Cumulative Sum Scaling, used in metagenomeSeq), "RLE" (Relative Log Expression, used in DESeq2), "GMPR" (Geometric Mean of Pairwise Ratio, introduced in Chen et al., 2018), Wrench (introduced in Kumar et al., 2018) or "none". Alternatively the user can supply its own vector or matrix of offsets (see note for specification of the user-supplied offsets).
...	Additional parameters passed on to specific methods (for now CSS and RLE)

Details

RLE has additional pseudocounts and type arguments to add pseudocounts to the observed counts (defaults to 0L) and to compute offsets using only positive counts (if type == "poscounts"). This mimics the behavior of `DESeq2::DESeq()` when using `sfType == "poscounts"`. CSS has an additional reference argument to choose the location function used to compute the reference quantiles (defaults to median as in the Nature publication but can be set to mean to reproduce behavior of functions `cumNormStat*` from metagenomeSeq). Wrench has two additional parameters: `groups` to specify sample groups and `type` to either reproduce exactly the default `Wrench::wrench()` behavior (type = "wrench", default) or to use simpler heuristics (type = "simple"). Note that (i) CSS normalization fails when the median absolute deviation around quantiles does not become instable for high quantiles (limited count variations both within and across samples) and/or one sample has less than two positive counts, (ii) RLE fails when there are no common species across all samples (unless type == "poscounts" has been specified) and (iii) GMPR fails if a sample does not share any species with all other samples.

Value

If `offset = "none"`, NULL else a vector of length `nrow(counts)` with one offset per sample.

References

- Chen, L., Reeve, J., Zhang, L., Huang, S., Wang, X. and Chen, J. (2018) GMPR: A robust normalization method for zero-inflated count data with application to microbiome sequencing data. *PeerJ*, 6, e4600 doi: [10.7717/peerj.4600](https://doi.org/10.7717/peerj.4600)
- Paulson, J. N., Colin Stine, O., Bravo, H. C. and Pop, M. (2013) Differential abundance analysis for microbial marker-gene surveys. *Nature Methods*, 10, 1200-1202 doi: [10.1038/nmeth.2658](https://doi.org/10.1038/nmeth.2658)
- Anders, S. and Huber, W. (2010) Differential expression analysis for sequence count data. *Genome Biology*, 11, R106 doi: [10.1186/gb20101110r106](https://doi.org/10.1186/gb20101110r106)
- Kumar, M., Slud, E., Okrah, K. et al. (2018) Analysis and correction of compositional bias in sparse sequencing count data. *BMC Genomics* 19, 799 doi: [10.1186/s1286401851605](https://doi.org/10.1186/s1286401851605)

Examples

```
data(trichoptera)
```

```

counts <- trichoptera$Abundance
compute_offset(counts)
## Other normalization schemes
compute_offset(counts, offset = "RLE", pseudocounts = 1)
compute_offset(counts, offset = "Wrench", groups = trichoptera$Covariate$Group)
compute_offset(counts, offset = "GMPR")
## User supplied offsets
my_offset <- setNames(rep(1, nrow(counts)), rownames(counts))
compute_offset(counts, offset = my_offset)

```

extract_probs

Extract edge selection frequency in bootstrap subsamples

Description

Extracts edge selection frequency in networks reconstructed from bootstrap subsamples during the stars stability selection procedure, as either a matrix or a named vector. In the latter case, edge names follow igraph naming convention.

Usage

```

extract_probs(
  Robject,
  penalty = NULL,
  index = NULL,
  crit = c("StARS", "BIC", "EBIC"),
  format = c("matrix", "vector"),
  tol = 1e-05
)

```

Arguments

Robject	an object with class <code>PLNnetworkfamily</code> , i.e. an output from <code>PLNnetwork()</code>
penalty	penalty used for the bootstrap subsamples
index	Integer index of the model to be returned. Only the first value is taken into account.
crit	a character for the criterion used to performed the selection. Either "BIC", "ICL", "EBIC", "StARS", "R_squared". Default is ICL for PLNPCA, and BIC for PLNnetwork. If StARS (Stability Approach to Regularization Selection) is chosen and stability selection was not yet performed, the function will call the method <code>stability_selection()</code> with default argument.
format	output format. Either a matrix (default) or a named vector.
tol	tolerance for rounding error when comparing penalties.

Value

Either a matrix or named vector of edge-wise probabilities. In the latter case, edge names follow igraph convention.

Examples

```

data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
nets <- PLNnetwork(Abundance ~ 1 + offset(log(Offset)), data = trichoptera)
## Not run:
stability_selection(nets)
probs <- extract_probs(nets, crit = "StARS", format = "vector")
probs

## End(Not run)

## Not run:
## Add edge attributes to graph using igraph
net_stars <- getBestModel(nets, "StARS")
g <- plot(net_stars, type = "partial_cor", plot=F)
library(igraph)
E(g)$prob <- probs[as_ids(E(g))]
g

## End(Not run)

```

fisher

Fisher information matrix for Theta

Description

Extracts Fisher information matrix of Θ from objects returned by [PLN](#) and its variants. Fisher matrix is computed using one of two approximation scheme: wald (default, conservative, gives large confidence interval) or louis (anticonservative). Note that the Fisher information matrix is the full-data version (scaled by the number of observations), usually noted

$$I_n(\theta)$$

Usage

```

fisher(object, type)

## S3 method for class 'PLNfit'
fisher(object, type = c("wald", "louis"))

```

Arguments

object	an R6 object with class PLNfit
type	Either wald (default) or louis. Approximation scheme used to compute the Fisher information matrix

Value

A block-diagonal matrix with p (number of species) blocks of size d (number of covariates), assuming Θ is a matrix of size $d * p$.

Methods (by class)

- `PLNfit`: Fisher information matrix for `PLNfit`

See Also

[standard_error](#) for standard errors

<code>fitted.PLNfit</code>	<i>Extracts model fitted values from objects returned by PLN() and its variants</i>
----------------------------	---

Description

Extracts model fitted values from objects returned by [PLN\(\)](#) and its variants

Usage

```
## S3 method for class 'PLNfit'
fitted(object, ...)
```

Arguments

<code>object</code>	an R6 object with class PLNfit
<code>...</code>	additional parameters for S3 compatibility. Not used

Value

A matrix of Fitted values extracted from the object `object`.

fitted.PLNmixturefit *Extracts model fitted values from objects returned by `PLNmixture()` and its variants*

Description

Extracts model fitted values from objects returned by `PLNmixture()` and its variants

Usage

```
## S3 method for class 'PLNmixturefit'
fitted(object, ...)
```

Arguments

object an R6 object with class `PLNmixturefit`
 ... additional parameters for S3 compatibility. Not used

Value

A matrix of Fitted values extracted from the object object.

getBestModel.PLNPCAfamily
Best model extraction from a collection of models

Description

Best model extraction from a collection of models

Usage

```
## S3 method for class 'PLNPCAfamily'
getBestModel(Robject, crit = c("ICL", "BIC"), ...)

getBestModel(Robject, crit, ...)

## S3 method for class 'PLNmixturefamily'
getBestModel(Robject, crit = c("ICL", "BIC"), ...)

## S3 method for class 'PLNnetworkfamily'
getBestModel(Robject, crit = c("BIC", "EBIC", "StARS"), ...)
```

Arguments

Robjct	an object with class PLNPCAfamily or PLNnetworkfamily
crit	a character for the criterion used to performed the selection. Either "BIC", "ICL", "EBIC", "StARS", "R_squared". Default is ICL for PLNPCA, and BIC for PLNnetwork. If StARS (Stability Approach to Regularization Selection) is chosen and stability selection was not yet performed, the function will call the method <code>stability_selection()</code> with default argument.
...	additional parameters for StARS criterion (only for PLNnetwork). <code>stability</code> , a scalar indicating the target stability ($= 1 - 2 \beta$) at which the network is selected. Default is 0.9.

Value

Send back an object with class `PLNPCAfit` or `PLNnetworkfit`

Methods (by class)

- PLNPCAfamily: Model extraction for `PLNPCAfamily`
- PLNmixturefamily: Model extraction for `PLNmixturefamily`
- PLNnetworkfamily: Model extraction for `PLNnetworkfamily`

Examples

```
## Not run:
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCA <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:4)
myModel <- getBestModel(myPCA)

## End(Not run)
```

`getModel.PLNPCAfamily` *Model extraction from a collection of models*

Description

Model extraction from a collection of models

Usage

```
## S3 method for class 'PLNPCAfamily'
getModel(Robjct, var, index = NULL)

getModel(Robjct, var, index)

## S3 method for class 'PLNmixturefamily'
```

```
getModel(Robject, var, index = NULL)

## S3 method for class 'PLNnetworkfamily'
getModel(Robject, var, index = NULL)
```

Arguments

Robject	an R6 object with class PLNPCAfamily or PLNnetworkfamily
var	value of the parameter (rank for PLNPCA , sparsity for PLNnetwork) that identifies the model to be extracted from the collection. If no exact match is found, the model with closest parameter value is returned with a warning.
index	Integer index of the model to be returned. Only the first value is taken into account.

Value

Sends back an object with class [PLNPCAfit](#) or [PLNnetworkfit](#).

Methods (by class)

- [PLNPCAfamily](#): Model extraction for [PLNPCAfamily](#)
- [PLNmixturefamily](#): Model extraction for [PLNmixturefamily](#)
- [PLNnetworkfamily](#): Model extraction for [PLNnetworkfamily](#)

Examples

```
## Not run:
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCA <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
myModel <- getModel(myPCA, 2)

## End(Not run)
```

mollusk

Mollusk data set

Description

This data set gives the abundance of 32 mollusk species in 163 samples. For each sample, 4 additional covariates are known.

Usage

```
mollusk
```

Format

A list with 2 two data frames:

Abundance a 163 x 32 data frame of abundancies/counts (163 samples and 32 mollusk species)

Covariate a 163 x 4 data frame of covariates:

site a factor with 8 levels indicating the sampling site

season a factor with 4 levels indicating the season

method a factor with 2 levels for the method of sampling - wood or string

duration a numeric with 3 levels for the time of exposure in week

In order to prepare the data for using formula in multivariate analysis (multiple outputs and inputs), use `prepare_data()`. Original data set has been extracted from ade4.

Source

Data from Richardot-Coulet, Chessel and Bournaud.

References

Richardot-Coulet, M., Chessel D. and Bournaud M. (1986) Typological value of the benthos of old beds of a large river. Methodological approach. Archiv für Hydrobiologie, 107, 363–383.

See Also

`prepare_data()`

Examples

```
data(mollusk)
mollusc <- prepare_data(mollusk$Abundance, mollusk$Covariate)
```

oaks

Oaks amplicon data set

Description

This data set gives the abundance of 114 taxa (66 bacterial OTU, 48 fungal OTUs) in 116 samples. For each sample, 11 additional covariates are known.

Usage

oaks

Format

A data frame with 13 variables:

- Abundance: A 114 taxa by 116 samples count matrix
- Offset: A 114 taxa by 116 samples offset matrix
- Sample: Unique sample id
- tree: Tree status with respect to the pathogen (susceptible, intermediate or resistant)
- branch: Unique branch id in each tree (4 branches were sampled in each tree, with 10 leaves per branch)
- leafNO: Unique leaf id in each tree (40 leaves were sampled in each tree)
- distTObase: Distance of the sampled leaf to the base of the branch
- distTOTrunk: Distance of the sampled leaf to the base of the tree trunk
- distTOground: Distance of the sampled leaf to the base of the ground
- pmInfection: Powdery mildew infection, proportion of the upper leaf area displaying mildew symptoms
- orientation: Orientation of the branch (South-West SW or North-East NE)
- readsTOTfun: Total number of ITS1 reads for that leaf
- readsTOTbac: Total number of 16S reads for that leaf

Source

Data from B. Jakuschkin and coauthors.

References

Jakuschkin, B., Fievet, V., Schwaller, L. et al. Deciphering the Pathobiome: Intra- and Interkingdom Interactions Involving the Pathogen *Erysiphe alphitoides*. *Microb Ecol* 72, 870–880 (2016). doi: [10.1007/s002480160777x](https://doi.org/10.1007/s002480160777x)

See Also

[prepare_data\(\)](#)

Examples

```
data(oaks)
## Not run:
oaks_networks <- PLNnetwork(formula = Abundance ~ 1 + offset(log(Offset)), data = oaks)

## End(Not run)
```

PLN

*Poisson lognormal model***Description**

Fit the multivariate Poisson lognormal model with a variational algorithm. Use the (g)lm syntax for model specification (covariates, offsets, weights).

Usage

```
PLN(formula, data, subset, weights, control = list())
```

Arguments

formula	an object of class "formula": a symbolic description of the model to be fitted.
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in data, the variables are taken from <code>environment(formula)</code> , typically the environment from which PLN is called.
subset	an optional vector specifying a subset of observations to be used in the fitting process.
weights	an optional vector of observation weights to be used in the fitting process.
control	a list for controlling the optimization. See details.

Details

The parameter control is a list controlling the optimization with the following entries:

- "covariance" character setting the model for the covariance matrix. Either "full", "diagonal" or "spherical". Default is "full".
- "trace" integer for verbosity.
- "inception" Set up the initialization. By default, the model is initialized with a multivariate linear model applied on log-transformed data, and with the same formula as the one provided by the user. However, the user can provide a PLNfit (typically obtained from a previous fit), which sometimes speeds up the inference.
- "ftol_rel" stop when an optimization step changes the objective function by less than ftol multiplied by the absolute value of the parameter. Default is 1e-6 when $n < p$, 1e-8 otherwise.
- "ftol_abs" stop when an optimization step changes the objective function by less than ftol multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than xtol multiplied by the absolute value of the parameter. Default is 1e-4
- "xtol_abs" stop when an optimization step changes every parameters by less than xtol multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds maxeval. Default is 10000

- "maxtime" stop when the optimization time (in seconds) exceeds maxtime. Default is -1 (no restriction)
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".

Value

an R6 object with class `PLNfit`

See Also

The class `PLNfit`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1, data = trichoptera)
```

PLNfamily

An R6 Class to represent a collection of PLNfit

Description

super class for `PLNPCAfamily` and `PLNnetworkfamily`. The R6 class benefits from S3 methods such as `getBestModel()`, `getModel()` and `plot()`.

Details

The parameter `control` is a list controlling the optimization with the following entries:

- "covariance" character setting the model for the covariance matrix. Either "full", "diagonal" or "spherical". Default is "full".
- "trace" integer for verbosity.
- "inception" Set up the initialization. By default, the model is initialized with a multivariate linear model applied on log-transformed data, and with the same formula as the one provided by the user. However, the user can provide a `PLNfit` (typically obtained from a previous fit), which sometimes speeds up the inference.
- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is $1e-6$ when $n < p$, $1e-8$ otherwise.
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is $1e-4$

- "xtol_abs" stop when an optimization step changes every parameters by less than xtol multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds maxeval. Default is 10000
- "maxtime" stop when the optimization time (in seconds) exceeds maxtime. Default is -1 (no restriction)
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".

Public fields

responses the matrix of responses common to every models

covariates the matrix of covariates common to every models

offsets the matrix of offsets common to every models

weights the vector of observation weights

inception a [PLNfit](#) object, obtained when no sparsifying penalty is applied.

models a list of [PLNfit](#) object, one per penalty.

Active bindings

criteria a data frame with the values of some criteria (approximated log-likelihood, BIC, ICL, etc.) for the collection of models / fits BIC and ICL are defined so that they are on the same scale as the model log-likelihood, i.e. with the form, $\text{loglik} - 0.5 \text{ penalty}$

convergence sends back a data frame with some convergence diagnostics associated with the optimization process (method, optimal value, etc)

Methods

Public methods:

- [PLNfamily\\$new\(\)](#)
- [PLNfamily\\$postTreatment\(\)](#)
- [PLNfamily\\$getModel\(\)](#)
- [PLNfamily\\$plot\(\)](#)
- [PLNfamily\\$show\(\)](#)
- [PLNfamily\\$print\(\)](#)
- [PLNfamily\\$clone\(\)](#)

Method [new\(\)](#): Create a new [PLNfamily](#) object.

Usage:

```
PLNfamily$new(responses, covariates, offsets, weights, control)
```

Arguments:

responses the matrix of responses common to every models

covariates the matrix of covariates common to every models

offsets the matrix of offsets common to every models

`weights` the vector of observation weights
`control` a list for controlling the optimization. See details.
Returns: A new [PLNfamily](#) object

Method `postTreatment()`: Update fields after optimization

Usage:
`PLNfamily$postTreatment()`

Method `getModel()`: Extract a model from a collection of models

Usage:
`PLNfamily$getModel(var, index = NULL)`
Arguments:
`var` value of the parameter (rank for PLNPCA, sparsity for PLNnetwork) that identifies the model to be extracted from the collection. If no exact match is found, the model with closest parameter value is returned with a warning.
`index` Integer index of the model to be returned. Only the first value is taken into account.
Returns: A [PLNfit](#) object

Method `plot()`: Lineplot of selected criteria for all models in the collection

Usage:
`PLNfamily$plot(criteria, reverse)`
Arguments:
`criteria` A valid model selection criteria for the collection of models. Includes loglik, BIC (all), ICL (PLNPCA) and pen_loglik, EBIC (PLNnetwork)
`reverse` A logical indicating whether to plot the value of the criteria in the "natural" direction (loglik - penalty) or in the "reverse" direction (-2 loglik + penalty). Default to FALSE, i.e use the natural direction, on the same scale as the log-likelihood.
Returns: A [ggplot2](#) object

Method `show()`: User friendly print method

Usage:
`PLNfamily$show()`

Method `print()`: User friendly print method

Usage:
`PLNfamily$print()`

Method `clone()`: The objects of this class are cloneable with this method.

Usage:
`PLNfamily$clone(deep = FALSE)`
Arguments:
`deep` Whether to make a deep clone.

See Also

[getModel\(\)](#)

Description

The function `PLN()` fit a model which is an instance of a object with class `PLNfit`. Objects produced by the functions `PLNnetwork()`, `PLNPCA()`, `PLNmixture()` and `PLNLDA()` also enjoy the methods of `PLNfit()` by inheritance.

This class comes with a set of R6 methods, some of them being useful for the user and exported as S3 methods. See the documentation for `coef()`, `sigma()`, `predict()`, `vcov()` and `standard_error()`.

Fields are accessed via active binding and cannot be changed by the user.

Details

The parameter control is a list controlling the optimization with the following entries:

- "covariance" character setting the model for the covariance matrix. Either "full", "diagonal" or "spherical". Default is "full".
- "trace" integer for verbosity.
- "inception" Set up the initialization. By default, the model is initialized with a multivariate linear model applied on log-transformed data, and with the same formula as the one provided by the user. However, the user can provide a `PLNfit` (typically obtained from a previous fit), which sometimes speeds up the inference.
- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is $1e-6$ when $n < p$, $1e-8$ otherwise.
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is $1e-4$
- "xtol_abs" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds `maxeval`. Default is 10000
- "maxtime" stop when the optimization time (in seconds) exceeds `maxtime`. Default is -1 (no restriction)
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".

Active bindings

`n` number of samples

`q` number of dimensions of the latent space

p number of species
 d number of covariates
 model_par a list with the matrices of parameters found in the model (Theta, Sigma, plus some others depending on the variant)
 fisher Variational approximation of the Fisher Information matrix
 std_err Variational approximation of the variance-covariance matrix of model parameters estimates.
 var_par a list with two matrices, M and S2, which are the estimated parameters in the variational approximation
 latent a matrix: values of the latent vector (Z in the model)
 latent_pos a matrix: values of the latent position vector (Z) without covariates effects or offset
 fitted a matrix: fitted values of the observations (A in the model)
 nb_param number of parameters in the current PLN model
 vcov_model character: the model used for the covariance (either "spherical", "diagonal" or "full")
 optim_par a list with parameters useful for monitoring the optimization
 weights observational weights
 loglik (weighted) variational lower bound of the loglikelihood
 loglik_vec element-wise variational lower bound of the loglikelihood
 BIC variational lower bound of the BIC
 entropy Entropy of the variational distribution
 ICL variational lower bound of the ICL
 R_squared approximated goodness-of-fit criterion
 criteria a vector with loglik, BIC, ICL and number of parameters

Methods

Public methods:

- `PLNfit$update()`
- `PLNfit$new()`
- `PLNfit$optimize()`
- `PLNfit$VEstep()`
- `PLNfit$set_R2()`
- `PLNfit$compute_fisher()`
- `PLNfit$compute_standard_error()`
- `PLNfit$postTreatment()`
- `PLNfit$predict()`
- `PLNfit$show()`
- `PLNfit$print()`
- `PLNfit$clone()`

Method `update()`: Update a `PLNfit` object

Usage:

```
PLNfit$update(
  Theta = NA,
  Sigma = NA,
  M = NA,
  S2 = NA,
  Ji = NA,
  R2 = NA,
  Z = NA,
  A = NA,
  monitoring = NA
)
```

Arguments:

Theta matrix of regression matrix

Sigma variance-covariance matrix of the latent variables

M matrix of mean vectors for the variational approximation

S2 matrix of variance vectors for the variational approximation

Ji vector of variational lower bounds of the log-likelihoods (one value per sample)

R2 approximate R^2 goodness-of-fit criterion

Z matrix of latent vectors (includes covariates and offset effects)

A matrix of fitted values

monitoring a list with optimization monitoring quantities

Returns: Update the current [PLNfit](#) object

Method `new()`: Initialize a [PLNfit](#) model

Usage:

```
PLNfit$new(responses, covariates, offsets, weights, formula, xlevels, control)
```

Arguments:

responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)-class

covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)-class

offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)-class

weights an optional vector of observation weights to be used in the fitting process.

formula model formula used for fitting, extracted from the formula in the upper-level call

xlevels named listed of factor levels included in the models, extracted from the formula in the upper-level call and used for predictions.

control a list for controlling the optimization. See details.

Method `optimize()`: Call to the C++ optimizer and update of the relevant fields

Usage:

```
PLNfit$optimize(responses, covariates, offsets, weights, control)
```

Arguments:

responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in PLNfamily-class
covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in PLNfamily-class
offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in PLNfamily-class
weights an optional vector of observation weights to be used in the fitting process.
control a list for controlling the optimization. See details.

Method `VEstep()`: Result of one call to the VE step of the optimization procedure: optimal variational parameters (M, S) and corresponding log likelihood values for fixed model parameters (Sigma, Theta). Intended to position new data in the latent space.

Usage:

```
PLNfit$VEstep(covariates, offsets, responses, weights, control = list())
```

Arguments:

covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in PLNfamily-class
offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in PLNfamily-class
responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in PLNfamily-class
weights an optional vector of observation weights to be used in the fitting process.
control a list for controlling the optimization. See details.

Returns: A list with three components:

- the matrix M of variational means,
- the matrix S2 of variational variances
- the vector `log.lik` of (variational) log-likelihood of each new observation

Method `set_R2()`: Update R2 field after optimization

Usage:

```
PLNfit$set_R2(responses, covariates, offsets, weights, nullModel = NULL)
```

Arguments:

responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in PLNfamily-class
covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in PLNfamily-class
offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in PLNfamily-class
weights an optional vector of observation weights to be used in the fitting process.
nullModel null model used for approximate R2 computations. Defaults to a GLM model with same design matrix but not latent variable.

Method `compute_fisher()`: Safely compute the fisher information matrix (FIM)

Usage:

```
PLNfit$compute_fisher(type = c("wald", "louis"), X = NULL)
```

Arguments:

type approximation scheme to compute the fisher information matrix. Either wald (default) or louis. type = "louis" results in smaller confidence intervals.

X design matrix used to compute the FIM

Returns: a sparse matrix with sensible dimension names

Method compute_standard_error(): Compute univariate standard error for coefficients of Theta from the FIM

Usage:

```
PLNfit$compute_standard_error()
```

Returns: a matrix of standard deviations.

Method postTreatment(): Update R2, fisher and std_err fields after optimization

Usage:

```
PLNfit$postTreatment(
  responses,
  covariates,
  offsets,
  weights = rep(1, nrow(responses)),
  type = c("wald", "louis", "none"),
  nullModel = NULL
)
```

Arguments:

responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in PLNfamily-class

covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in PLNfamily-class

offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in PLNfamily-class

weights an optional vector of observation weights to be used in the fitting process.

type approximation scheme to compute the fisher information matrix. Either wald (default) or louis. type = "louis" results in smaller confidence intervals.

nullModel null model used for approximate R2 computations. Defaults to a GLM model with same design matrix but not latent variable.

Method predict(): Predict position, scores or observations of new data.

Usage:

```
PLNfit$predict(newdata, type = c("link", "response"), envir = parent.frame())
```

Arguments:

newdata A data frame in which to look for variables with which to predict. If omitted, the fitted values are used.

type Scale used for the prediction. Either link (default, predicted positions in the latent space) or response (predicted counts).

envir Environment in which the prediction is evaluated

Returns: A matrix with predictions scores or counts.

Method show(): User friendly print method

Usage:

```
PLNfit$show(
  model = paste("A multivariate Poisson Lognormal fit with", private$covariance,
    "covariance model.\n")
)
```

Arguments:

model First line of the print output

Method print(): User friendly print method

Usage:

```
PLNfit$print()
```

Method clone(): The objects of this class are cloneable with this method.

Usage:

```
PLNfit$clone(deep = FALSE)
```

Arguments:

deep Whether to make a deep clone.

Examples

```
## Not run:
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1, data = trichoptera)
class(myPLN)
print(myPLN)

## End(Not run)
```

Description

Fit the Poisson lognormal for LDA with a variational algorithm. Use the (g)lm syntax for model specification (covariates, offsets).

Usage

```
PLNLDA(formula, data, subset, weights, grouping, control = list())
```

Arguments

formula	an object of class "formula": a symbolic description of the model to be fitted.
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in data, the variables are taken from <code>environment(formula)</code> , typically the environment from which <code>lm</code> is called.
subset	an optional vector specifying a subset of observations to be used in the fitting process.
weights	an optional vector of observation weights to be used in the fitting process.
grouping	a factor specifying the class of each observation used for discriminant analysis.
control	a list for controlling the optimization process. See details.

Details

The parameter `control` is a list controlling the optimization with the following entries:

- "covariance" character setting the model for the covariance matrix. Either "full" or "spherical". Default is "full".
- "trace" integer for verbosity.
- "inception" Set up the initialization. By default, the model is initialized with a multivariate linear model applied on log-transformed data. However, the user can provide a `PLNfit` (typically obtained from a previous fit), which often speed up the inference.
- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is $1e-8$
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is $1e-4$
- "xtol_abs" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds `maxeval`. Default is 10000
- "maxtime" stop when the optimization time (in seconds) exceeds `maxtime`. Default is -1 (no restriction)
- "algorithm" the optimization method used by `NLOPT` among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See `NLOPT` documentation for further details. Default is "CCSAQ".

Value

an R6 object with class `PLNLDAfit()`

See Also

The class `PLNLDAfit`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLNLDA <- PLNLDA(Abundance ~ 1, grouping = Group, data = trichoptera)
```

 PLNLDAfit

An R6 Class to represent a PLNfit in a LDA framework

Description

The function `PLNLDA()` produces an instance of an object with class `PLNLDAfit`.

This class comes with a set of methods, some of them being useful for the user: See the documentation for the methods inherited by `PLNfit()`, the `plot()` method for LDA visualization and `predict()` method for prediction

Super class

`PLNmodels::PLNfit -> PLNLDAfit`

Active bindings

`rank` the dimension of the current model

`nb_param` number of parameters in the current PLN model

`model_par` a list with the matrices associated with the estimated parameters of the PLN model: Theta (covariates), Sigma (latent covariance), B (latent loadings), P (latent position) and Mu (group means)

`percent_var` the percent of variance explained by each axis

`corr_map` a matrix of correlations to plot the correlation circles

`scores` a matrix of scores to plot the individual factor maps

`group_means` a matrix of group mean vectors in the latent space.

Methods**Public methods:**

- `PLNLDAfit$new()`
- `PLNLDAfit$optimize()`
- `PLNLDAfit$postTreatment()`
- `PLNLDAfit$setVisualization()`
- `PLNLDAfit$plot_individual_map()`
- `PLNLDAfit$plot_correlation_map()`
- `PLNLDAfit$plot_LDA()`
- `PLNLDAfit$predict()`
- `PLNLDAfit$show()`

- `PLNLDAfit$clone()`

Method `new()`: Initialize a `PLNLDAfit` object

Usage:

```
PLNLDAfit$new(
  grouping,
  responses,
  covariates,
  offsets,
  weights,
  formula,
  xlevels,
  control
)
```

Arguments:

`grouping` a factor specifying the class of each observation used for discriminant analysis.

`responses` the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in `PLNfamily-class`

`covariates` design matrix (called X in the model). Will usually be extracted from the corresponding field in `PLNfamily-class`

`offsets` offset matrix (called O in the model). Will usually be extracted from the corresponding field in `PLNfamily-class`

`weights` an optional vector of observation weights to be used in the fitting process.

`formula` model formula used for fitting, extracted from the formula in the upper-level call

`xlevels` named listed of factor levels included in the models, extracted from the formula in the upper-level call and used for predictions.

`control` a list for controlling the optimization. See details.

Method `optimize()`: Compute group means and axis of the LDA (noted B in the model) in the latent space, update corresponding fields

Usage:

```
PLNLDAfit$optimize(grouping, covariates, control)
```

Arguments:

`grouping` design matrix for the grouping variable

`covariates` design matrix. Automatically built from the covariates and the formula from the call

`control` a list for controlling the optimization. See details.

`X` Abundance matrix.

Method `postTreatment()`: Update R2, fisher and `std_err` fields and visualization after optimization

Usage:

```
PLNLDAfit$postTreatment(grouping, responses, covariates, offsets)
```

Arguments:

grouping a factor specifying the class of each observation used for discriminant analysis.
 responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in PLNfamily-class
 covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in PLNfamily-class
 offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in PLNfamily-class

Method `setVisualization()`: Compute LDA scores in the latent space and update corresponding fields.

Usage:

```
PLNLDAfit$setVisualization(scale.unit = FALSE)
```

Arguments:

`scale.unit` Logical. Should LDA scores be rescaled to have unit variance

Method `plot_individual_map()`: Plot the factorial map of the LDA

Usage:

```
PLNLDAfit$plot_individual_map(
  axes = 1:min(2, self$rank),
  main = "Individual Factor Map",
  plot = TRUE
)
```

Arguments:

`axes` numeric, the axes to use for the plot when `map = "individual" or "variable"`. Default is `c(1,min(rank))`

`main` character. A title for the single plot (individual or variable factor map). If NULL (the default), an hopefully appropriate title will be used.

`plot` logical. Should the plot be displayed or sent back as ggplot object

Returns: a [ggplot](#) graphic

Method `plot_correlation_map()`: Plot the correlation circle of a specified axis for a [PLNLDAfit](#) object

Usage:

```
PLNLDAfit$plot_correlation_map(
  axes = 1:min(2, self$rank),
  main = "Variable Factor Map",
  cols = "default",
  plot = TRUE
)
```

Arguments:

`axes` numeric, the axes to use for the plot when `map = "individual" or "variable"`. Default is `c(1,min(rank))`

`main` character. A title for the single plot (individual or variable factor map). If NULL (the default), an hopefully appropriate title will be used.

`cols` a character, factor or numeric to define the color associated with the variables. By default, all variables receive the default color of the current palette.

`plot` logical. Should the plot be displayed or sent back as ggplot object

Returns: a `ggplot` graphic

Method `plot_LDA()`: Plot a summary of the `PLNLDAfit` object

Usage:

```
PLNLDAfit$plot_LDA(
  nb_axes = min(3, self$rank),
  var_cols = "default",
  plot = TRUE
)
```

Arguments:

`nb_axes` scalar: the number of axes to be considered when `map = "both"`. The default is `min(3,rank)`.

`var_cols` a character, factor or numeric to define the color associated with the variables. By default, all variables receive the default color of the current palette.

`plot` logical. Should the plot be displayed or sent back as ggplot object

Returns: a `grob` object

Method `predict()`: Predict group of new samples

Usage:

```
PLNLDAfit$predict(
  newdata,
  type = c("posterior", "response", "scores"),
  scale = c("log", "prob"),
  prior = NULL,
  control = list(),
  envir = parent.frame()
)
```

Arguments:

`newdata` A data frame in which to look for variables, offsets and counts with which to predict.

`type` The type of prediction required. The default are posterior probabilities for each group (in either unnormalized log-scale or natural probabilities, see "scale" for details), "response" is the group with maximal posterior probability and "scores" is the average score along each separation axis in the latent space, with weights equal to the posterior probabilities.

`scale` The scale used for the posterior probability. Either log-scale ("log", default) or natural probabilities summing up to 1 ("prob").

`prior` User-specified prior group probabilities in the new data. If NULL (default), prior probabilities are computed from the learning set.

`control` a list for controlling the optimization. See `PLN()` for details.

`envir` Environment in which the prediction is evaluated

Method `show()`: User friendly print method

Usage:

```
PLNLDAfit$show()
```

Method `clone()`: The objects of this class are cloneable with this method.

Usage:

```
PLNLDAfit$clone(deep = FALSE)
```

Arguments:

`deep` Whether to make a deep clone.

See Also

The function [PLNLDA](#).

Examples

```
## Not run:
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLNLDA <- PLNLDA(Abundance ~ 1, grouping = Group, data = trichoptera)
class(myPLNLDA)
print(myPLNLDA)

## End(Not run)
```

PLNmixture

Poisson lognormal mixture model

Description

Fit the mixture variants of the Poisson lognormal with a variational algorithm. Use the (g)lm syntax for model specification (covariates, offsets).

Usage

```
PLNmixture(
  formula,
  data,
  subset,
  clusters = 1:5,
  control_init = list(),
  control_main = list()
)
```

Arguments

<code>formula</code>	an object of class "formula": a symbolic description of the model to be fitted.
<code>data</code>	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in data, the variables are taken from <code>environment(formula)</code> , typically the environment from which <code>lm</code> is called.
<code>subset</code>	an optional vector specifying a subset of observations to be used in the fitting process.
<code>clusters</code>	a vector of integer containing the successive number of clusters (or components) to be considered
<code>control_init</code>	a list for controlling the optimization at initialization. See details.
<code>control_main</code>	a list for controlling the main optimization process. See details.

Details

The list of parameters `control_init` and `control_main` control the optimization of the initialization and the main process, with the following entries

- "covariance" character setting the model for the covariance matrices of the mixture components. Either "full", "diagonal" or "spherical". Default is "spherical".
- "trace" integer for verbosity.
- "inception" Set up the initialization. By default, the model is initialized with a multivariate linear model applied on log-transformed data, and with the same formula as the one provided by the user. However, the user can provide a PLNfit (typically obtained from a previous fit), which sometimes speeds up the inference.
- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is $1e-6$ when $n < p$, $1e-8$ otherwise.
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is $1e-4$
- "xtol_abs" stop when an optimization step changes every parameters by less than `xtol` multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds `maxeval`. Default is 10000
- "maxtime" stop when the optimization time (in seconds) exceeds `maxtime`. Default is -1 (no restriction)
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".
- "ftol_out" outer solver stops when an optimization step changes the objective function by less than `xtol` multiply by the absolute value of the parameter. Default is $1e-6$
- "maxit_out" outer solver stops when the number of iteration exceeds `out.maxit`. Default is 50
- "smoothing" The smoothing to apply. Either, 'forward', 'backward' or 'both'. Default is 'both'.
- "iterates" number of forward/backward iteration of smoothing. Default is 2.

Value

an R6 object with class `PLNmixturefamily`, which contains a collection of models with class `PLNmixturefit`

See Also

The classes `PLNmixturefamily` and `PLNmixturefit`

Examples

```
## Use future to dispatch the computations on 2 workers
## Not run:
future::plan("multisession", workers = 2)

## End(Not run)

data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myMixtures <- PLNmixture(Abundance ~ 1 + offset(log(Offset)), data = trichoptera,
  control_main = list(smoothing = "forward", iterates = 1))

# Shut down parallel workers
## Not run:
future::plan("sequential")

## End(Not run)
```

PLNmixturefamily	<i>An R6 Class to represent a collection of PLNmixturefit</i>
------------------	---

Description

The function `PLNmixture()` produces an instance of this class.

This class comes with a set of methods, some of them being useful for the user: See the documentation for `getBestModel()`, `getModel()` and `plot()`.

Super class

`PLNmodels::PLNfamily` -> `PLNmixturefamily`

Active bindings

`clusters` vector indicating the number of clusters considered is the successively fitted models

Methods

Public methods:

- `PLNmixturefamily$new()`
- `PLNmixturefamily$optimize()`
- `PLNmixturefamily$smooth()`
- `PLNmixturefamily$plot()`
- `PLNmixturefamily$plot_objective()`
- `PLNmixturefamily$getBestModel()`
- `PLNmixturefamily$show()`
- `PLNmixturefamily$print()`
- `PLNmixturefamily$clone()`

Method `new()`: Initialize all models in the collection.

Usage:

```
PLNmixturefamily$new(
  clusters,
  responses,
  covariates,
  offsets,
  formula,
  xlevels,
  control
)
```

Arguments:

`clusters` the dimensions of the successively fitted models

`responses` the matrix of responses common to every models

`covariates` the matrix of covariates common to every models

`offsets` the matrix of offsets common to every models

`formula` model formula used for fitting, extracted from the formula in the upper-level call

`xlevels` named listed of factor levels included in the models, extracted from the formula in the upper-level call #'

`control` a list for controlling the optimization. See details.

`control` a list for controlling the optimization. See details.

Method `optimize()`: Call to the optimizer on all models of the collection

Usage:

```
PLNmixturefamily$optimize(control)
```

Arguments:

`control` a list for controlling the optimization. See details.

`control` a list for controlling the optimization. See details.

Method `smooth()`: function to restart clustering to avoid local minima by smoothing the log-likelihood values as a function of the number of clusters

Usage:

```
PLNmixturefamily$smooth(control)
```

Arguments:

control a list to control the smoothing process

Method plot(): Lineplot of selected criteria for all models in the collection

Usage:

```
PLNmixturefamily$plot(criteria = c("loglik", "BIC", "ICL"), reverse = FALSE)
```

Arguments:

criteria A valid model selection criteria for the collection of models. Any of "loglik", "BIC" or "ICL" (all).

reverse A logical indicating whether to plot the value of the criteria in the "natural" direction (loglik - 0.5 penalty) or in the "reverse" direction (-2 loglik + penalty). Default to FALSE, i.e use the natural direction, on the same scale as the log-likelihood..

Returns: A [ggplot2](#) object

Method plot_objective(): Plot objective value of the optimization problem along the penalty path

Usage:

```
PLNmixturefamily$plot_objective()
```

Returns: a [ggplot](#) graph

Method getBestModel(): Extract best model in the collection

Usage:

```
PLNmixturefamily$getBestModel(crit = c("BIC", "ICL", "loglik"))
```

Arguments:

crit a character for the criterion used to performed the selection. Either "BIC", "ICL" or "loglik". Default is ICL

Returns: a [PLNmixturefit](#) object

Method show(): User friendly print method

Usage:

```
PLNmixturefamily$show()
```

Method print(): User friendly print method

Usage:

```
PLNmixturefamily$print()
```

Method clone(): The objects of this class are cloneable with this method.

Usage:

```
PLNmixturefamily$clone(deep = FALSE)
```

Arguments:

deep Whether to make a deep clone.

See Also

The function [PLNmixture](#), the class [PLNmixturefit](#)

 PLNmixturefit

An R6 Class to represent a PLNfit in a mixture framework

Description

The function `PLNmixture` produces a collection of models which are instances of object with class `PLNmixturefit`. A `PLNmixturefit` (say, with k components) is itself a collection of k `PLNfit`.

This class comes with a set of methods, some of them being useful for the user: See the documentation for ...

Active bindings

`n` number of samples
`p` number of dimensions of the latent space
`k` number of components
`d` number of covariates
`components` components of the mixture (PLNfits)
`latent` a matrix: values of the latent vector (Z in the model)
`latent_pos` a matrix: values of the latent position vector (Z) without covariates effects or offset
`posteriorProb` matrix of posterior probability for cluster belonging
`memberships` vector for cluster index
`mixtureParam` vector of cluster proportions
`optim_par` a list with parameters useful for monitoring the optimization
`nb_param` number of parameters in the current PLN model
`entropy_clustering` Entropy of the variational distribution of the cluster (multinomial)
`entropy_latent` Entropy of the variational distribution of the latent vector (Gaussian)
`entropy` Full entropy of the variational distribution (latent vector + clustering)
`loglik` variational lower bound of the loglikelihood
`loglik_vec` element-wise variational lower bound of the loglikelihood
`BIC` variational lower bound of the BIC
`ICL` variational lower bound of the ICL (include entropy of both the clustering and latent distributions)
`R_squared` approximated goodness-of-fit criterion
`criteria` a vector with loglik, BIC, ICL, and number of parameters
`model_par` a list with the matrices of parameters found in the model (Theta, Sigma, Mu and Pi)
`vcov_model` character: the model used for the covariance (either "spherical", "diagonal" or "full")
`fitted` a matrix: fitted values of the observations (A in the model)
`group_means` a matrix of group mean vectors in the latent space.

Methods

Public methods:

- `PLNmixturefit$new()`
- `PLNmixturefit$optimize()`
- `PLNmixturefit$predict()`
- `PLNmixturefit$plot_clustering_data()`
- `PLNmixturefit$plot_clustering_pca()`
- `PLNmixturefit$postTreatment()`
- `PLNmixturefit$show()`
- `PLNmixturefit$print()`
- `PLNmixturefit$clone()`

Method `new()`: Optimize a the
Initialize a `PLNmixturefit` model

Usage:

```
PLNmixturefit$new(
  responses,
  covariates,
  offsets,
  posteriorProb,
  formula,
  xlevels,
  control
)
```

Arguments:

`responses` the matrix of responses common to every models
`covariates` the matrix of covariates common to every models
`offsets` the matrix of offsets common to every models
`posteriorProb` matrix of posterior probability for cluster belonging
`formula` model formula used for fitting, extracted from the formula in the upper-level call
`xlevels` named listed of factor levels included in the models, extracted from the formula in the upper-level call #'
`control` a list for controlling the optimization. See details.
`control` a list for controlling the optimization. See details.

Method `optimize()`: Optimize a `PLNmixturefit` model

Usage:

```
PLNmixturefit$optimize(responses, covariates, offsets, control)
```

Arguments:

`responses` the matrix of responses common to every models
`covariates` the matrix of covariates common to every models
`offsets` the matrix of offsets common to every models
`control` a list for controlling the optimization. See details.

`control` a list for controlling the optimization. See details.

Method `predict()`: Predict group of new samples

Usage:

```
PLNmixturefit$predict(
  newdata,
  type = c("posterior", "response", "position"),
  prior = matrix(rep(1/self$k, self$k), nrow(newdata), self$k, byrow = TRUE),
  control = list(),
  envir = parent.frame()
)
```

Arguments:

`newdata` A data frame in which to look for variables, offsets and counts with which to predict.

`type` The type of prediction required. The default posterior are posterior probabilities for each group, `response` is the group with maximal posterior probability and `latent` is the averaged latent coordinate (without offset and nor covariate effects), with weights equal to the posterior probabilities.

`prior` User-specified prior group probabilities in the new data. The default uses a uniform prior.

`control` a list for controlling the optimization. See `PLN()` for details.

`envir` Environment in which the prediction is evaluated

Method `plot_clustering_data()`: Plot the matrix of expected mean counts (without offsets, without covariate effects) reordered according the inferred clustering

Usage:

```
PLNmixturefit$plot_clustering_data(
  main = "Expected counts reorder by clustering",
  plot = TRUE,
  log_scale = TRUE
)
```

Arguments:

`main` character. A title for the plot. An hopefully appropriate title will be used by default.

`plot` logical. Should the plot be displayed or sent back as `ggplot` object

`log_scale` logical. Should the color scale values be log-transform before plotting? Default is TRUE.

Returns: a `ggplot` graphic

Method `plot_clustering_pca()`: Plot the individual map of a PCA performed on the latent coordinates, where individuals are colored according to the memberships

Usage:

```
PLNmixturefit$plot_clustering_pca(
  main = "Clustering labels in Individual Factor Map",
  plot = TRUE
)
```

Arguments:

`main` character. A title for the plot. An hopefully appropriate title will be used by default.

`plot` logical. Should the plot be displayed or sent back as [ggplot](#) object

Returns: a [ggplot](#) graphic

Method `postTreatment()`: Update fields after optimization

Usage:

```
PLNmixturefit$postTreatment(responses, covariates, offsets, weights, nullModel)
```

Arguments:

`responses` the matrix of responses common to every models

`covariates` the matrix of covariates common to every models

`offsets` the matrix of offsets common to every models

`weights` an optional vector of observation weights to be used in the fitting process.

`nullModel` null model used for approximate R2 computations. Defaults to a GLM model with same design matrix but not latent variable.

Method `show()`: User friendly print method

Usage:

```
PLNmixturefit$show()
```

Method `print()`: User friendly print method

Usage:

```
PLNmixturefit$print()
```

Method `clone()`: The objects of this class are cloneable with this method.

Usage:

```
PLNmixturefit$clone(deep = FALSE)
```

Arguments:

`deep` Whether to make a deep clone.

See Also

The function [PLNmixture](#), the class [PLNmixturefamily](#)

Description

The Poisson lognormal model and variants can be used for a variety of multivariate problems when count data are at play (including PCA or LDA for count data, network inference). This package implements efficient variational algorithms to fit such models accompanied with a set of functions for visualization and diagnostic.

Multivariate Poisson lognormal model (aka PLN)

See the main function [PLN\(\)](#) and the associated methods for manipulation.

Also try `vignette("PLN_trichoptera", package="PLNmodels")` for an overview.

Rank Constrained Poisson lognormal for Poisson Principal Component Analysis (aka PLNPCA)

See the main function [PLNPCA\(\)](#) and the associated methods for manipulation.

The Poisson PCA and the associated variational inference is fully explained in Chiquet et al (2018), see reference below.

Also try `vignette("PLNPCA_trichoptera", package="PLNmodels")` for an overview.

Sparse Poisson lognormal model for sparse covariance inference for counts (aka PLNnetwork)

See the main function [PLNnetwork\(\)](#) and the associated methods for manipulation.

Also try `vignette("PLNnetwork_trichoptera", package="PLNmodels")` for an overview.

Poisson lognormal discriminant analysis (aka PLNLDA)

See the main function [PLNLDA\(\)](#) and the associated methods for manipulation.

Also try `vignette("PLNLDA_trichoptera", package="PLNmodels")` for an overview.

Mixtures of Poisson lognormal models for model-based clustering (aka PLNmixture)

See the main function [PLNmixture\(\)](#) and the associated methods for manipulation.

Also try `vignette("PLNmixture_trichoptera", package="PLNmodels")` for an overview.

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Description

Fit the sparse inverse covariance variant of the Poisson lognormal with a variational algorithm. Use the (g)lm syntax for model specification (covariates, offsets).

Usage

```

PLNnetwork(
  formula,
  data,
  subset,
  weights,
  penalties = NULL,
  control_init = list(),
  control_main = list()
)

```

Arguments

formula	an object of class "formula": a symbolic description of the model to be fitted.
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in data, the variables are taken from <code>environment(formula)</code> , typically the environment from which <code>lm</code> is called.
subset	an optional vector specifying a subset of observations to be used in the fitting process.
weights	an optional vector of observation weights to be used in the fitting process.
penalties	an optional vector of positive real number controlling the level of sparsity of the underlying network. if <code>NULL</code> (the default), will be set internally. See <code>control_init</code> and <code>control_main</code> options for additional tuning of the penalty.
control_init	a list for controlling the optimization of the PLN model used at initialization, and how the vector of <code>penalties</code> is generated. See details.
control_main	a list for controlling the main optimization process. Can be used to specify adaptive penalty weights. See details.

Details

The list of parameters `control_main` controls the optimization of the main process, with the following entries:

- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is $1e-6$ when $n < p$, $1e-8$ otherwise.
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is 0
- "xtol_rel" stop when an optimization step changes every parameters by less than `xtol_rel` multiplied by the absolute value of the parameter. Default is $1e-4$
- "xtol_abs" stop when an optimization step changes every parameters by less than `xtol_abs`. Default is 0
- "maxeval" stop when the number of iteration exceeds `maxeval`. Default is 10000
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".

- "trace" integer for verbosity. Useless when cores > 1
- "ftol_out" outer solver stops when an optimization step changes the objective function by less than xtol multiply by the absolute value of the parameter. Default is 1e-6
- "maxit_out" outer solver stops when the number of iteration exceeds out.maxit. Default is 50
- "penalize_diagonal" boolean: should the diagonal terms be penalized in the graphical-Lasso? Default is TRUE
- "penalty_weights" p x p matrix of weights (default filled with 1) to adapt the amount of shrinkage to each pairs of node. Must be symmetric with positive values.

The list of parameters `control_init` controls the optimization process in the initialization and in the function `PLN()`, plus two additional parameters:

- "nPenalties" an integer that specified the number of values for the penalty grid when internally generated. Ignored when penalties is non NULL
- "min.ratio" the penalty grid ranges from the minimal value that produces a sparse to this value multiplied by min.ratio. Default is 0.1.

Value

an R6 object with class `PLNnetworkfamily`, which contains a collection of models with class `PLNnetworkfit`

See Also

The classes `PLNnetworkfamily` and `PLNnetworkfit`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
```

PLNnetworkfamily

An R6 Class to represent a collection of PLNnetworkfit

Description

The function `PLNnetwork()` produces an instance of this class.

This class comes with a set of methods, some of them being useful for the user: See the documentation for `getBestModel()`, `getModel()` and `plot()`

Super class

`PLNmodels::PLNfamily -> PLNnetworkfamily`

Active bindings

penalties the sparsity level of the network in the successively fitted models
 stability_path the stability path of each edge as returned by the stars procedure
 stability mean edge stability along the penalty path
 criteria a data frame with the values of some criteria (approximated log-likelihood, (E)BIC, ICL and R2, stability) for the collection of models / fits BIC, ICL and EBIC are defined so that they are on the same scale as the model log-likelihood, i.e. with the form, $\text{loglik} - 0.5 \text{ penalty}$

Methods**Public methods:**

- `PLNnetworkfamily$new()`
- `PLNnetworkfamily$optimize()`
- `PLNnetworkfamily$stability_selection()`
- `PLNnetworkfamily$coefficient_path()`
- `PLNnetworkfamily$getBestModel()`
- `PLNnetworkfamily$plot()`
- `PLNnetworkfamily$plot_stars()`
- `PLNnetworkfamily$plot_objective()`
- `PLNnetworkfamily$show()`
- `PLNnetworkfamily$clone()`

Method `new()`: Initialize all models in the collection

Usage:

```
PLNnetworkfamily$new(
  penalties,
  responses,
  covariates,
  offsets,
  weights,
  formula,
  xlevels,
  control
)
```

Arguments:

penalties a vector of positive real number controlling the level of sparsity of the underlying network.
 responses the matrix of responses common to every models
 covariates the matrix of covariates common to every models
 offsets the matrix of offsets common to every models
 weights the vector of observation weights
 formula model formula used for fitting, extracted from the formula in the upper-level call
 xlevels named listed of factor levels included in the models, extracted from the formula in the upper-level call and used for predictions.

`control` a list for controlling the optimization. See details.

Returns: Update current [PLNnetworkfit](#) with smart starting values

Method `optimize()`: Call to the C++ optimizer on all models of the collection

Usage:

```
PLNnetworkfamily$optimize(control)
```

Arguments:

`control` a list for controlling the optimization. See details.

Method `stability_selection()`: Compute the stability path by stability selection

Usage:

```
PLNnetworkfamily$stability_selection(subsamples = NULL, control = list())
```

Arguments:

`subsamples` a list of vectors describing the subsamples. The number of vectors (or list length) determines the number of subsamples used in the stability selection. Automatically set to 20 subsamples with size $10 \cdot \sqrt{n}$ if $n \geq 144$ and $0.8 \cdot n$ otherwise following Liu et al. (2010) recommendations.

`control` a list controlling the main optimization process in each call to `PLNnetwork`. See [PLNnetwork\(\)](#) for details.

Method `coefficient_path()`: Extract the regularization path of a [PLNnetworkfamily](#)

Usage:

```
PLNnetworkfamily$coefficient_path(precision = TRUE, corr = TRUE)
```

Arguments:

`precision` Logical. Should the regularization path be extracted from the precision matrix Omega (TRUE, default) or from the variance matrix Sigma (FALSE)

`corr` Logical. Should the matrix be transformed to (partial) correlation matrix before extraction? Defaults to TRUE

Method `getBestModel()`: Extract the best network in the family according to some criteria

Usage:

```
PLNnetworkfamily$getBestModel(
  crit = c("BIC", "EBIC", "StARS"),
  stability = 0.9
)
```

Arguments:

`crit` character. Criterion used to perform the selection. Is "StARS" is chosen but `$stability` field is empty, will compute stability path.

`stability` Only used for "StARS" criterion. A scalar indicating the target stability (= $1 - 2 \beta$) at which the network is selected. Default is 0.9.

Method `plot()`: Display various outputs (goodness-of-fit criteria, robustness, diagnostic) associated with a collection of `PLNnetwork` fits (a [PLNnetworkfamily](#))

Usage:

```

PLNnetworkfamily$plot(
  criteria = c("loglik", "pen_loglik", "BIC", "EBIC"),
  reverse = FALSE,
  log.x = TRUE
)

```

Arguments:

`criteria` vector of characters. The criteria to plot in `c("loglik", "pen_loglik", "BIC", "EBIC")`.

Defaults to all of them.

`reverse` A logical indicating whether to plot the value of the criteria in the "natural" direction (`loglik - 0.5 penalty`) or in the "reverse" direction (`-2 loglik + penalty`). Default to `FALSE`, i.e use the natural direction, on the same scale as the log-likelihood..

`log.x` logical: should the x-axis be represented in log-scale? Default is `TRUE`.

Returns: a `ggplot` graph

Method `plot_stars()`: Plot stability path*Usage:*

```
PLNnetworkfamily$plot_stars(stability = 0.9, log.x = TRUE)
```

Arguments:

`stability` scalar: the targeted level of stability in stability plot. Default is `0.9`.

`log.x` logical: should the x-axis be represented in log-scale? Default is `TRUE`.

Returns: a `ggplot` graph

Method `plot_objective()`: Plot objective value of the optimization problem along the penalty path*Usage:*

```
PLNnetworkfamily$plot_objective()
```

Returns: a `ggplot` graph

Method `show()`: User friendly print method*Usage:*

```
PLNnetworkfamily$show()
```

Method `clone()`: The objects of this class are cloneable with this method.*Usage:*

```
PLNnetworkfamily$clone(deep = FALSE)
```

Arguments:

`deep` Whether to make a deep clone.

See Also

The function `PLNnetwork()`, the class `PLNnetworkfit`

Examples

```

data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
class(fits)

```

PLNnetworkfit	<i>An R6 Class to represent a PLNfit in a sparse inverse covariance framework</i>
---------------	---

Description

The function `PLNnetworkfit()` produces a collection of models which are instances of object with class `PLNnetworkfit`.

This class comes with a set of methods, some of them being useful for the user: See the documentation for `plot()` and methods inherited from `PLNfit`.

Super class

`PLNmodels::PLNfit -> PLNnetworkfit`

Active bindings

`penalty` the level of sparsity in the current model

`n_edges` number of edges if the network (non null coefficient of the sparse precision matrix)

`nb_param` number of parameters in the current PLN model

`pen_loglik` variational lower bound of the l1-penalized loglikelihood

`model_par` a list with the matrices associated with the estimated parameters of the pPCA model: Theta (covariates), Sigma (latent covariance) and Theta (latent precision matrix). Note Omega and Sigma are inverse of each other.

`EBIC` variational lower bound of the EBIC

`density` proportion of non-null edges in the network

`criteria` a vector with loglik, penalized loglik, BIC, EBIC, ICL, R_squared, number of parameters, number of edges, and graph density

Methods

Public methods:

- `PLNnetworkfit$new()`
- `PLNnetworkfit$update()`
- `PLNnetworkfit$optimize()`
- `PLNnetworkfit$postTreatment()`
- `PLNnetworkfit$latent_network()`
- `PLNnetworkfit$plot_network()`
- `PLNnetworkfit$show()`
- `PLNnetworkfit$clone()`

Method `new()`: Initialize a `PLNnetworkfit` object

Usage:

```

PLNnetworkfit$new(
  penalty,
  responses,
  covariates,
  offsets,
  weights,
  formula,
  xlevels,
  control
)

```

Arguments:

penalty a positive real number controlling the level of sparsity of the underlying network.

responses the matrix of responses common to every models

covariates the matrix of covariates common to every models

offsets the matrix of offsets common to every models

weights an optional vector of observation weights to be used in the fitting process.

formula model formula used for fitting, extracted from the formula in the upper-level call

xlevels named listed of factor levels included in the models, extracted from the formula in [PLNnetwork\(\)](#) call

control a list for controlling the optimization of the PLN model used at initialization. See [PLNnetwork\(\)](#) for details.

Method `update()`: Update fields of a `PLNnetworkfit` object

Usage:

```

PLNnetworkfit$update(
  penalty = NA,
  Theta = NA,
  Sigma = NA,
  Omega = NA,
  M = NA,
  S2 = NA,
  Z = NA,
  A = NA,
  Ji = NA,
  R2 = NA,
  monitoring = NA
)

```

Arguments:

penalty a positive real number controlling the level of sparsity of the underlying network.

Theta matrix of regression matrix

Sigma variance-covariance matrix of the latent variables

Omega precision matrix of the latent variables. Inverse of *Sigma*.

M matrix of mean vectors for the variational approximation

S2 matrix of variance vectors for the variational approximation

Z matrix of latent vectors (includes covariates and offset effects)

A matrix of fitted values
 Ji vector of variational lower bounds of the log-likelihoods (one value per sample)
 R2 approximate R² goodness-of-fit criterion
 monitoring a list with optimization monitoring quantities

Method optimize(): Call to the C++ optimizer and update of the relevant fields

Usage:

```
PLNnetworkfit$optimize(responses, covariates, offsets, weights, control)
```

Arguments:

responses the matrix of responses common to every models
 covariates the matrix of covariates common to every models
 offsets the matrix of offsets common to every models
 weights an optional vector of observation weights to be used in the fitting process.
 control a list for controlling the optimization of the PLN model used at initialization. See [PLNnetwork\(\)](#) for details.

Method postTreatment(): Compute PCA scores in the latent space and update corresponding fields.

Usage:

```
PLNnetworkfit$postTreatment(responses, covariates, offsets, weights, nullModel)
```

Arguments:

responses the matrix of responses common to every models
 covariates the matrix of covariates common to every models
 offsets the matrix of offsets common to every models
 weights an optional vector of observation weights to be used in the fitting process.
 nullModel null model used for approximate R2 computations. Defaults to a GLM model with same design matrix but not latent variable.

Method latent_network(): Extract interaction network in the latent space

Usage:

```
PLNnetworkfit$latent_network(type = c("partial_cor", "support", "precision"))
```

Arguments:

type edge value in the network. Can be "support" (binary edges), "precision" (coefficient of the precision matrix) or "partial_cor" (partial correlation between species)

Returns: a square matrix of size PLNnetworkfit\$n

Method plot_network(): plot the latent network.

Usage:

```
PLNnetworkfit$plot_network(
  type = c("partial_cor", "support"),
  output = c("igraph", "corrplot"),
  edge.color = c("#F8766D", "#00BFC4"),
  remove.isolated = FALSE,
  node.labels = NULL,
```



```

    layout = layout_in_circle,
    plot = TRUE
  )

```

Arguments:

type edge value in the network. Either "precision" (coefficient of the precision matrix) or "partial_cor" (partial correlation between species).

output Output type. Either igraph (for the network) or corrplot (for the adjacency matrix)

edge.color Length 2 color vector. Color for positive/negative edges. Default is c("#F8766D", "#00BFC4"). Only relevant for igraph output.

remove.isolated if TRUE, isolated node are remove before plotting. Only relevant for igraph output.

node.labels vector of character. The labels of the nodes. The default will use the column names of the response matrix.

layout an optional igraph layout. Only relevant for igraph output.

plot logical. Should the final network be displayed or only sent back to the user. Default is TRUE.

Method show(): User friendly print method

Usage:

```
PLNnetworkfit$show()
```

Method clone(): The objects of this class are cloneable with this method.

Usage:

```
PLNnetworkfit$clone(deep = FALSE)
```

Arguments:

deep Whether to make a deep clone.

See Also

The function [PLNnetwork\(\)](#), the class [PLNnetworkfamily](#)

Examples

```

## Not run:
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
nets <- PLNnetwork(Abundance ~ 1, data = trichoptera)
myPLNnet <- getBestModel(nets)
class(myPLNnet)
print(myPLNnet)

## End(Not run)

```

Description

Fit the PCA variants of the Poisson lognormal with a variational algorithm. Use the (g)lm syntax for model specification (covariates, offsets).

Usage

```
PLNPCA(
  formula,
  data,
  subset,
  weights,
  ranks = 1:5,
  control_init = list(),
  control_main = list()
)
```

Arguments

formula	an object of class "formula": a symbolic description of the model to be fitted.
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in data, the variables are taken from <code>environment(formula)</code> , typically the environment from which <code>lm</code> is called.
subset	an optional vector specifying a subset of observations to be used in the fitting process.
weights	an optional vector of observation weights to be used in the fitting process.
ranks	a vector of integer containing the successive ranks (or number of axes to be considered)
control_init	a list for controlling the optimization at initialization. See details of function PLN() .
control_main	a list for controlling the main optimization process. See details.

Details

The list of parameters `control_main` controls the optimization of the main process, with the following entries:

- "ftol_rel" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is `1e-8`
- "ftol_abs" stop when an optimization step changes the objective function by less than `ftol` multiplied by the absolute value of the parameter. Default is `0`

- "xtol_rel" stop when an optimization step changes every parameters by less than xtol multiplied by the absolute value of the parameter. Default is 1e-4
- "xtol_abs" stop when an optimization step changes every parameters by less than xtol multiplied by the absolute value of the parameter. Default is 0
- "maxeval" stop when the number of iteration exceeds maxeval. Default is 10000
- "maxtime" stop when the optimization time (in seconds) exceeds maxtime. Default is -1 (no restriction)
- "algorithm" the optimization method used by NLOPT among LD type, i.e. "CCSAQ", "MMA", "LBFGS", "VAR1", "VAR2". See NLOPT documentation for further details. Default is "CCSAQ".
- "trace" integer for verbosity.

Value

an R6 object with class `PLNPCAfamily`, which contains a collection of models with class `PLNPCAfit`

See Also

The classes `PLNPCAfamily` and `PLNPCAfit`

Examples

```

#' ## Use future to dispatch the computations on 2 workers
## Not run:
future::plan("multisession", workers = 2)

## End(Not run)

data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCA <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)

# Shut down parallel workers
## Not run:
future::plan("sequential")

## End(Not run)

```

PLNPCAfamily

An R6 Class to represent a collection of PLNPCAfit

Description

The function `PLNPCA()` produces an instance of this class.

This class comes with a set of methods, some of them being useful for the user: See the documentation for `getBestModel()`, `getModel()` and `plot()`.

Super class

PLNmodels::PLNfamily -> PLNPCAfamily

Active bindings

ranks the dimensions of the successively fitted models

Methods**Public methods:**

- PLNPCAfamily\$new()
- PLNPCAfamily\$optimize()
- PLNPCAfamily\$getModel()
- PLNPCAfamily\$getBestModel()
- PLNPCAfamily\$plot()
- PLNPCAfamily\$show()
- PLNPCAfamily\$clone()

Method new(): Initialize all models in the collection.

Usage:

```
PLNPCAfamily$new(
  ranks,
  responses,
  covariates,
  offsets,
  weights,
  formula,
  xlevels,
  control
)
```

Arguments:

ranks the dimensions of the successively fitted models

responses the matrix of responses common to every models

covariates the matrix of covariates common to every models

offsets the matrix of offsets common to every models

weights the vector of observation weights

formula model formula used for fitting, extracted from the formula in the upper-level call

xlevels named listed of factor levels included in the models, extracted from the formula in the upper-level call and used for predictions.

control a list for controlling the optimization. See details.

Method optimize(): Call to the C++ optimizer on all models of the collection

Usage:

```
PLNPCAfamily$optimize(control)
```

Arguments:

control a list for controlling the optimization. See details.

Method `getModel()`: Extract model from collection and add "PCA" class for compatibility with `factoextra::fviz()`

Usage:

```
PLNPCAfamily$getModel(var, index = NULL)
```

Arguments:

var value of the parameter (rank for PLNPCA, sparsity for PLNnetwork) that identifies the model to be extracted from the collection. If no exact match is found, the model with closest parameter value is returned with a warning.

index Integer index of the model to be returned. Only the first value is taken into account.

Returns: a `PLNPCAfit` object

Method `getBestModel()`: Extract best model in the collection

Usage:

```
PLNPCAfamily$getBestModel(crit = c("ICL", "BIC"))
```

Arguments:

crit a character for the criterion used to performed the selection. Either "ICL", "BIC". Default is ICL

Returns: a `PLNPCAfit` object

Method `plot()`: Lineplot of selected criteria for all models in the collection

Usage:

```
PLNPCAfamily$plot(criteria = c("loglik", "BIC", "ICL"), reverse = FALSE)
```

Arguments:

criteria A valid model selection criteria for the collection of models. Any of "loglik", "BIC" or "ICL" (all).

reverse A logical indicating whether to plot the value of the criteria in the "natural" direction (loglik - penalty) or in the "reverse" direction (-2 loglik + penalty). Default to FALSE, i.e use the natural direction, on the same scale as the log-likelihood.

Returns: A `ggplot2` object

Method `show()`: User friendly print method

Usage:

```
PLNPCAfamily$show()
```

Method `clone()`: The objects of this class are cloneable with this method.

Usage:

```
PLNPCAfamily$clone(deep = FALSE)
```

Arguments:

deep Whether to make a deep clone.

See Also

The function `PLNPCA()`, the class `PLNPCAfit()`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCAs <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
class(myPCAs)
```

 PLNPCAfit

An R6 Class to represent a PLNfit in a PCA framework

Description

The function `PLNPCA()` produces a collection of models which are instances of object with class `PLNPCAfit`. This class comes with a set of methods, some of them being useful for the user: See the documentation for the methods inherited by `PLNfit` and the `plot()` methods for PCA visualization

Super class

`PLNmodels::PLNfit` -> `PLNPCAfit`

Active bindings

`rank` the dimension of the current model

`nb_param` number of parameters in the current PLN model

`entropy` entropy of the variational distribution

`latent_pos` a matrix: values of the latent position vector (Z) without covariates effects or offset

`model_par` a list with the matrices associated with the estimated parameters of the pPCA model: Theta (covariates), Sigma (latent covariance) and B (latent loadings)

`percent_var` the percent of variance explained by each axis

`corr_circle` a matrix of correlations to plot the correlation circles

`scores` a matrix of scores to plot the individual factor maps (a.k.a. principal components)

`rotation` a matrix of rotation of the latent space

`eig` description of the eigenvalues, similar to `percent_var` but for use with external methods

`var` a list of data frames with PCA results for the variables: `coord` (coordinates of the variables), `cor` (correlation between variables and dimensions), `cos2` (Cosine of the variables) and `contrib` (contributions of the variable to the axes)

`ind` a list of data frames with PCA results for the individuals: `coord` (coordinates of the individuals), `cos2` (Cosine of the individuals), `contrib` (contributions of individuals to an axis inertia) and `dist` (distance of individuals to the origin).

`call` Hacky binding for compatibility with `factoextra` functions

Methods

Public methods:

- `PLNPCAfit$new()`
- `PLNPCAfit$update()`
- `PLNPCAfit$optimize()`
- `PLNPCAfit$setVisualization()`
- `PLNPCAfit$postTreatment()`
- `PLNPCAfit$compute_fisher()`
- `PLNPCAfit$plot_individual_map()`
- `PLNPCAfit$plot_correlation_circle()`
- `PLNPCAfit$plot_PCA()`
- `PLNPCAfit$show()`
- `PLNPCAfit$clone()`

Method `new()`: Initialize a `PLNPCAfit` object

Usage:

```
PLNPCAfit$new(
  rank,
  responses,
  covariates,
  offsets,
  weights,
  formula,
  xlevels,
  control
)
```

Arguments:

`rank` rank of the PCA (or equivalently, dimension of the latent space)

`responses` the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`covariates` design matrix (called X in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`offsets` offset matrix (called O in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`weights` an optional vector of observation weights to be used in the fitting process.

`formula` model formula used for fitting, extracted from the formula in the upper-level call

`xlevels` named listed of factor levels included in the models, extracted from the formula in the upper-level call and used for predictions.

`control` a list for controlling the optimization. See details.

Method `update()`: Update a `PLNPCAfit` object

Usage:

```

PLNPCAfit$update(
  Theta = NA,
  Sigma = NA,
  B = NA,
  M = NA,
  S2 = NA,
  Z = NA,
  A = NA,
  Ji = NA,
  R2 = NA,
  monitoring = NA
)

```

Arguments:

Theta matrix of regression matrix

Sigma variance-covariance matrix of the latent variables

B matrix of PCA loadings (in the latent space)

M matrix of mean vectors for the variational approximation

S2 matrix of variance vectors for the variational approximation

Z matrix of latent vectors (includes covariates and offset effects)

A matrix of fitted values

Ji vector of variational lower bounds of the log-likelihoods (one value per sample)

R2 approximate R^2 goodness-of-fit criterion

monitoring a list with optimization monitoring quantities

Returns: Update the current [PLNPCAfit](#) object

Method `optimize()`: Call to the C++ optimizer and update of the relevant fields

Usage:

```
PLNPCAfit$optimize(responses, covariates, offsets, weights, control)
```

Arguments:

responses the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)

covariates design matrix (called X in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)

offsets offset matrix (called O in the model). Will usually be extracted from the corresponding field in [PLNfamily](#)

weights an optional vector of observation weights to be used in the fitting process.

control a list for controlling the optimization. See details.

Method `setVisualization()`: Compute PCA scores in the latent space and update corresponding fields.

Usage:

```
PLNPCAfit$setVisualization(scale.unit = FALSE)
```

Arguments:

scale.unit Logical. Should PCA scores be rescaled to have unit variance

Method `postTreatment()`: Update R2, fisher, std_err fields and set up visualization after optimization

Usage:

```
PLNPCAfit$postTreatment(responses, covariates, offsets, weights, nullModel)
```

Arguments:

`responses` the matrix of responses (called Y in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`covariates` design matrix (called X in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`offsets` offset matrix (called O in the model). Will usually be extracted from the corresponding field in `PLNfamily`

`weights` an optional vector of observation weights to be used in the fitting process.

`nullModel` null model used for approximate R2 computations. Defaults to a GLM model with same design matrix but not latent variable.

Method `compute_fisher()`: Safely compute the fisher information matrix (FIM)

Usage:

```
PLNPCAfit$compute_fisher(type = c("wald", "louis"), X = NULL)
```

Arguments:

`type` approximation scheme to compute the fisher information matrix. Either `wald` (default) or `louis`. `type = "louis"` results in smaller confidence intervals.

`X` design matrix used to compute the FIM

Returns: a sparse matrix with sensible dimension names

Method `plot_individual_map()`: Plot the factorial map of the PCA

Usage:

```
PLNPCAfit$plot_individual_map(
  axes = 1:min(2, self$rank),
  main = "Individual Factor Map",
  plot = TRUE,
  cols = "default"
)
```

Arguments:

`axes` numeric, the axes to use for the plot when `map = "individual"` or `"variable"`. Default is `c(1,min(rank))`

`main` character. A title for the single plot (individual or variable factor map). If `NULL` (the default), an hopefully appropriate title will be used.

`plot` logical. Should the plot be displayed or sent back as `ggplot` object

`cols` a character, factor or numeric to define the color associated with the individuals. By default, all individuals receive the default color of the current palette.

Returns: a `ggplot` graphic

Method `plot_correlation_circle()`: Plot the correlation circle of a specified axis for a `PLNLDAfit` object

Usage:

```
PLNPCAfit$plot_correlation_circle(
  axes = 1:min(2, self$rank),
  main = "Variable Factor Map",
  cols = "default",
  plot = TRUE
)
```

Arguments:

axes numeric, the axes to use for the plot when *map* = "individual" or "variable". Default is `c(1,min(rank))`

main character. A title for the single plot (individual or variable factor map). If NULL (the default), an hopefully appropriate title will be used.

cols a character, factor or numeric to define the color associated with the variables. By default, all variables receive the default color of the current palette.

plot logical. Should the plot be displayed or sent back as ggplot object

Returns: a [ggplot](#) graphic

Method `plot_PCA()`: Plot a summary of the [PLNPCAfit](#) object

Usage:

```
PLNPCAfit$plot_PCA(
  nb_axes = min(3, self$rank),
  ind_cols = "ind_cols",
  var_cols = "var_cols",
  plot = TRUE
)
```

Arguments:

nb_axes scalar: the number of axes to be considered when *map* = "both". The default is `min(3,rank)`.

ind_cols a character, factor or numeric to define the color associated with the individuals. By default, all variables receive the default color of the current palette.

var_cols a character, factor or numeric to define the color associated with the variables. By default, all variables receive the default color of the current palette.

plot logical. Should the plot be displayed or sent back as ggplot object

Returns: a [grob](#) object

Method `show()`: User friendly print method

Usage:

```
PLNPCAfit$show()
```

Method `clone()`: The objects of this class are cloneable with this method.

Usage:

```
PLNPCAfit$clone(deep = FALSE)
```

Arguments:

deep Whether to make a deep clone.

See Also

The function [PLNPCA](#), the class [PLNPCAfamily](#)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCAs <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
myPCA <- getBestModel(myPCAs)
class(myPCA)
print(myPCA)
```

plot.PLNfamily	<i>Display the criteria associated with a collection of PLN fits (a PLNfamily)</i>
----------------	--

Description

Display the criteria associated with a collection of PLN fits (a PLNfamily)

Usage

```
## S3 method for class 'PLNfamily'
plot(x, criteria = c("loglik", "BIC", "ICL"), reverse = FALSE, ...)
```

Arguments

x	an R6 object with class PLNfamily
criteria	vector of characters. The criteria to plot in c("loglik", "BIC", "ICL"). Default is c("loglik", "BIC", "ICL").
reverse	A logical indicating whether to plot the value of the criteria in the "natural" direction (loglik - 0.5 penalty) or in the "reverse" direction (-2 loglik + penalty). Default to FALSE, i.e use the natural direction, on the same scale as the log-likelihood.
...	additional parameters for S3 compatibility. Not used

Details

The BIC and ICL criteria have the form 'loglik - 1/2 * penalty' so that they are on the same scale as the model log-likelihood. You can change this direction and use the alternate form '-2*loglik + penalty', as some authors do, by setting reverse = TRUE.

Value

Produces a plot representing the evolution of the criteria of the different models considered, highlighting the best model in terms of BIC and ICL (see details).

See Also

[plot.PLNPCAfamily\(\)](#) and [plot.PLNnetworkfamily\(\)](#)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCAs <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
## Not run:
plot(myPCAs)

## End(Not run)
```

plot.PLNLDAfit	<i>LDA visualization (individual and/or variable factor map(s)) for a PLNPCAfit object</i>
----------------	--

Description

LDA visualization (individual and/or variable factor map(s)) for a [PLNPCAfit](#) object

Usage

```
## S3 method for class 'PLNLDAfit'
plot(
  x,
  map = c("both", "individual", "variable"),
  nb_axes = min(3, x$rank),
  axes = seq.int(min(2, x$rank)),
  var_cols = "var_colors",
  plot = TRUE,
  main = NULL,
  ...
)
```

Arguments

x	an R6 object with class <code>PLNPCAfit</code>
map	the type of output for the PCA visualization: either "individual", "variable" or "both". Default is "both".
nb_axes	scalar: the number of axes to be considered when map = "both". The default is min(3,rank).
axes	numeric, the axes to use for the plot when map = "individual" or "variable". Default is c(1,min(rank))
var_cols	a character or factor to define the color associated with the variables. By default, all variables receive the default color of the current palette.

plot	logical. Should the plot be displayed or sent back as <code>ggplot2</code> object
main	character. A title for the single plot (individual or variable factor map). If NULL (the default), an hopefully appropriate title will be used.
...	Not used (S3 compatibility).

Value

displays an individual and/or variable factor maps for the corresponding axes, and/or sends back a `ggplot2` or `gtable` object

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLNLDA <- PLNLDA(Abundance ~ 1, grouping = Group, data = trichoptera)
## Not run:
plot(myPLNLDA, map = "individual", nb_axes = 2)

## End(Not run)
```

plot.PLNmixturefamily *Display the criteria associated with a collection of PLNmixture fits (a PLNmixturefamily)*

Description

Display the criteria associated with a collection of PLNmixture fits (a PLNmixturefamily)

Usage

```
## S3 method for class 'PLNmixturefamily'
plot(
  x,
  type = c("criteria", "diagnostic"),
  criteria = c("loglik", "BIC", "ICL"),
  reverse = FALSE,
  ...
)
```

Arguments

x	an R6 object with class <code>PLNmixturefamily</code>
type	a character, either "criteria" or "diagnostic" for the type of plot.
criteria	vector of characters. The criteria to plot in <code>c("loglik", "BIC", "ICL")</code> . Default is <code>c("loglik", "BIC", "ICL")</code> .

reverse	A logical indicating whether to plot the value of the criteria in the "natural" direction (loglik - 0.5 penalty) or in the "reverse" direction (-2 loglik + penalty). Default to FALSE, i.e use the natural direction, on the same scale as the log-likelihood.
...	additional parameters for S3 compatibility. Not used

Details

The BIC and ICL criteria have the form 'loglik - 1/2 * penalty' so that they are on the same scale as the model log-likelihood. You can change this direction and use the alternate form '-2*loglik + penalty', as some authors do, by setting reverse = TRUE.

Value

Produces either a diagnostic plot (with type = 'diagnostic') or the evolution of the criteria of the different models considered (with type = 'criteria', the default).

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myMixtures <- PLNmixture(Abundance ~ 1 + offset(log(Offset)),
  data = trichoptera, control_main = list(smoothing = "forward", iterates = 1))
plot(myMixtures, reverse = TRUE)
```

plot.PLNmixturefit *Mixture visualization of a PLNmixturefit object*

Description

Represent the result of the clustering either by coloring the individual in a two-dimension PCA factor map, or by representing the expected matrix of count reorder according to the clustering.

Usage

```
## S3 method for class 'PLNmixturefit'
plot(x, type = c("pca", "matrix"), main = NULL, plot = TRUE, ...)
```

Arguments

x	an R6 object with class PLNmixturefit
type	character for the type of plot, either "pca", for or "matrix". Default is "pca".
main	character. A title for the plot. If NULL (the default), an hopefully appropriate title will be used.
plot	logical. Should the plot be displayed or sent back as ggplot object
...	Not used (S3 compatibility).

Value

a `ggplot` graphic

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLNmixture(Abundance ~ 1 + offset(log(Offset)),
  data = trichoptera, control_main = list(iterates = 0)) %>% getBestModel()
plot(myPLN, "pca")
plot(myPLN, "matrix")
```

`plot.PLNnetworkfamily` *Display various outputs (goodness-of-fit criteria, robustness, diagnostic) associated with a collection of PLNnetwork fits (a [PLNnetworkfamily](#))*

Description

Display various outputs (goodness-of-fit criteria, robustness, diagnostic) associated with a collection of PLNnetwork fits (a [PLNnetworkfamily](#))

Usage

```
## S3 method for class 'PLNnetworkfamily'
plot(
  x,
  type = c("criteria", "stability", "diagnostic"),
  criteria = c("loglik", "pen_loglik", "BIC", "EBIC"),
  reverse = FALSE,
  log.x = TRUE,
  stability = 0.9,
  ...
)
```

Arguments

<code>x</code>	an R6 object with class PLNnetworkfamily
<code>type</code>	a character, either "criteria", "stability" or "diagnostic" for the type of plot.
<code>criteria</code>	vector of characters. The criteria to plot in <code>c("loglik", "BIC", "ICL", "R_squared", "EBIC", "pen_loglik")</code> . Default is <code>c("loglik", "pen_loglik", "BIC", "EBIC")</code> . Only relevant when <code>type = "criteria"</code> .
<code>reverse</code>	A logical indicating whether to plot the value of the criteria in the "natural" direction (<code>loglik - 0.5 penalty</code>) or in the "reverse" direction (<code>-2 loglik + penalty</code>). Default to <code>FALSE</code> , i.e use the natural direction, on the same scale as the log-likelihood.

log.x logical: should the x-axis be represented in log-scale? Default is TRUE.
 stability scalar: the targeted level of stability in stability plot. Default is .9.
 ... additional parameters for S3 compatibility. Not used

Details

The BIC and ICL criteria have the form 'loglik - 1/2 * penalty' so that they are on the same scale as the model log-likelihood. You can change this direction and use the alternate form '-2*loglik + penalty', as some authors do, by setting reverse = TRUE.

Value

Produces either a diagnostic plot (with type = 'diagnostic'), a stability plot (with type = 'stability') or the evolution of the criteria of the different models considered (with type = 'criteria', the default).

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
## Not run:
plot(fits)

## End(Not run)
```

plot.PLNnetworkfit *Extract and plot the network (partial correlation, support or inverse covariance) from a [PLNnetworkfit](#) object*

Description

Extract and plot the network (partial correlation, support or inverse covariance) from a [PLNnetworkfit](#) object

Usage

```
## S3 method for class 'PLNnetworkfit'
plot(
  x,
  type = c("partial_cor", "support"),
  output = c("igraph", "corrplot"),
  edge.color = c("#F8766D", "#00BFC4"),
  remove.isolated = FALSE,
  node.labels = NULL,
  layout = layout_in_circle,
  plot = TRUE,
  ...
)
```


Arguments

x	an R6 object with class <code>PLNnetworkfit</code>
type	character. Value of the weight of the edges in the network, either "partial_cor" (partial correlation) or "support" (binary). Default is "partial_cor".
output	the type of output used: either 'igraph' or 'corrplot'. Default is 'igraph'.
edge.color	Length 2 color vector. Color for positive/negative edges. Default is <code>c("#F8766D", "#00BFC4")</code> . Only relevant for igraph output.
remove.isolated	if TRUE, isolated node are remove before plotting. Only relevant for igraph output.
node.labels	vector of character. The labels of the nodes. The default will use the column names of the response matrix.
layout	an optional igraph layout. Only relevant for igraph output.
plot	logical. Should the final network be displayed or only sent back to the user. Default is TRUE.
...	Not used (S3 compatibility).

Value

Send back an invisible object (igraph or Matrix, depending on the output chosen) and optionally displays a graph (via igraph or corrplot for large ones)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
myNet <- getBestModel(fits)
## Not run:
plot(myNet)

## End(Not run)
```

plot.PLNPCAfamily	<i>Display the criteria associated with a collection of PLNPCA fits (a PLNPCAfamily)</i>
-------------------	--

Description

Display the criteria associated with a collection of PLNPCA fits (a PLNPCAfamily)

Usage

```
## S3 method for class 'PLNPCAfamily'
plot(x, criteria = c("loglik", "BIC", "ICL"), reverse = FALSE, ...)
```

Arguments

x	an R6 object with class <code>PLNPCAfamily</code>
criteria	vector of characters. The criteria to plot in <code>c("loglik", "BIC", "ICL")</code> . Default is <code>c("loglik", "BIC", "ICL")</code> .
reverse	A logical indicating whether to plot the value of the criteria in the "natural" direction (<code>loglik - 0.5 penalty</code>) or in the "reverse" direction (<code>-2 loglik + penalty</code>). Default to <code>FALSE</code> , i.e use the natural direction, on the same scale as the log-likelihood.
...	additional parameters for S3 compatibility. Not used

Details

The BIC and ICL criteria have the form '`loglik - 1/2 * penalty`' so that they are on the same scale as the model log-likelihood. You can change this direction and use the alternate form '`-2*loglik + penalty`', as some authors do, by setting `reverse = TRUE`.

Value

Produces a plot representing the evolution of the criteria of the different models considered, highlighting the best model in terms of BIC and ICL (see details).

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCAs <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
## Not run:
plot(myPCAs)

## End(Not run)
```

plot.PLNPCAfit	<i>PCA visualization (individual and/or variable factor map(s)) for a <code>PLNPCAfit</code> object</i>
----------------	---

Description

PCA visualization (individual and/or variable factor map(s)) for a `PLNPCAfit` object

Usage

```
## S3 method for class 'PLNPCAfit'
plot(
  x,
  map = c("both", "individual", "variable"),
  nb_axes = min(3, x$rank),
  axes = seq.int(min(2, x$rank)),
```

```

    ind_cols = "ind_colors",
    var_cols = "var_colors",
    plot = TRUE,
    main = NULL,
    ...
  )

```

Arguments

x	an R6 object with class PLNPCAfit
map	the type of output for the PCA visualization: either "individual", "variable" or "both". Default is "both".
nb_axes	scalar: the number of axes to be considered when map = "both". The default is min(3,rank).
axes	numeric, the axes to use for the plot when map = "individual" or map = "variable". Default is c(1,min(rank))
ind_cols	a character, factor or numeric to define the color associated with the individuals. By default, all variables receive the default color of the current palette.
var_cols	a character, factor or numeric to define the color associated with the variables. By default, all variables receive the default color of the current palette.
plot	logical. Should the plot be displayed or sent back as ggplot object
main	character. A title for the single plot (individual or variable factor map). If NULL (the default), an hopefully appropriate title will be used.
...	Not used (S3 compatibility).

Value

displays an individual and/or variable factor maps for the corresponding axes, and/or sends back a [ggplot](#) or [gtable](#) object

Examples

```

data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPCAs <- PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = trichoptera, ranks = 1:5)
myPCA <- getBestModel(myPCAs)
## Not run:
plot(myPCA, map = "individual", nb_axes=2, ind_cols = trichoptera$Group)
plot(myPCA, map = "variable", nb_axes=2)
plot(myPCA, map = "both", nb_axes=2, ind_cols = trichoptera$Group)

## End(Not run)

```

predict.PLNfit *Predict counts of a new sample*

Description

Predict counts of a new sample

Usage

```
## S3 method for class 'PLNfit'
predict(object, newdata, type = c("link", "response"), ...)
```

Arguments

object	an R6 object with class <code>PLNfit</code>
newdata	A data frame in which to look for variables and offsets with which to predict
type	The type of prediction required. The default is on the scale of the linear predictors (i.e. log average count)
...	additional parameters for S3 compatibility. Not used

Value

A matrix of predicted log-counts (if `type = "link"`) or predicted counts (if `type = "response"`).

predict.PLNLDAfit *Predict group of new samples*

Description

Predict group of new samples

Usage

```
## S3 method for class 'PLNLDAfit'
predict(
  object,
  newdata,
  type = c("posterior", "response", "scores"),
  scale = c("log", "prob"),
  prior = NULL,
  control = list(),
  ...
)
```

Arguments

object	an R6 object with class <code>PLNLDAfit</code>
newdata	A data frame in which to look for variables, offsets and counts with which to predict.
type	The type of prediction required. The default are posterior probabilities for each group (in either unnormalized log-scale or natural probabilities, see "scale" for details), "response" is the group with maximal posterior probability and "scores" is the average score along each separation axis in the latent space, with weights equal to the posterior probabilities.
scale	The scale used for the posterior probability. Either log-scale ("log", default) or natural probabilities summing up to 1 ("prob").
prior	User-specified prior group probabilities in the new data. If NULL (default), prior probabilities are computed from the learning set.
control	a list for controlling the optimization. See <code>PLN()</code> for details.
...	additional parameters for S3 compatibility. Not used

Value

A matrix of posterior probabilities for each group (if type = "posterior"), a matrix of (average) scores in the latent space (if type = "scores") or a vector of predicted groups (if type = "response").

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myLDA <- PLNLDA(Abundance ~ 0 + offset(log(Offset)),
                grouping = Group,
                data = trichoptera)

## Not run:
post_probs <- predict(myLDA, newdata = trichoptera, type = "posterior", scale = "prob")
head(round(post_probs, digits = 3))
predicted_group <- predict(myLDA, newdata = trichoptera, type = "response")
table(predicted_group, trichoptera$Group, dnn = c("predicted", "true"))

## End(Not run)
```

predict.PLNmixturefit *Prediction for a PLNmixturefit object*

Description

Predict either posterior probabilities for each group or latent positions based on new samples

Usage

```
## S3 method for class 'PLNmixturefit'
predict(
  object,
  newdata,
  type = c("posterior", "response", "position"),
  prior = matrix(rep(1/object$k, object$k), nrow(newdata), object$k, byrow = TRUE),
  control = list(),
  ...
)
```

Arguments

<code>object</code>	an R6 object with class <code>PLNmixturefit</code>
<code>newdata</code>	A data frame in which to look for variables, offsets and counts with which to predict.
<code>type</code>	The type of prediction required. The default posterior are posterior probabilities for each group, response is the group with maximal posterior probability and latent is the averaged latent in the latent space, with weights equal to the posterior probabilities.
<code>prior</code>	User-specified prior group probabilities in the new data. The default uses a uniform prior.
<code>control</code>	a list for controlling the optimization. See <code>PLN()</code> for details.
<code>...</code>	additional parameters for S3 compatibility. Not used

Value

A matrix of posterior probabilities for each group (if type = "posterior"), a matrix of (average) position in the latent space (if type = "position") or a vector of predicted groups (if type = "response").

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLNmixture(Abundance ~ 1 + offset(log(Offset)),
  data = trichoptera, control_main = list(iterates = 0)) %>% getBestModel()
predict(myPLN, trichoptera, "posterior")
predict(myPLN, trichoptera, "position")
predict(myPLN, trichoptera, "response")
```

prepare_data	<i>Prepare data for use in PLN models</i>
--------------	---

Description

Prepare data in proper format for use in PLN model and its variants. The function (i) merges a count table and a covariate data frame in the most comprehensive way and (ii) computes offsets from the count table using one of several normalization schemes (TSS, CSS, RLE, GMPR, Wrench, etc). The function fails with informative messages when the heuristics used for sample matching fail.

Usage

```
prepare_data(counts, covariates, offset = "TSS", ...)
```

Arguments

counts	Required. An abundance count table, preferably with dimensions names and species as columns.
covariates	Required. A covariates data frame, preferably with row names.
offset	Optional. Normalization scheme used to compute scaling factors used as offset during PLN inference. Available schemes are "TSS" (Total Sum Scaling, default), "CSS" (Cumulative Sum Scaling, used in metagenomeSeq), "RLE" (Relative Log Expression, used in DESeq2), "GMPR" (Geometric Mean of Pairwise Ratio, introduced in Chen et al., 2018), Wrench (introduced in Kumar et al., 2018) or "none". Alternatively the user can supply its own vector or matrix of offsets (see note for specification of the user-supplied offsets).
...	Additional parameters passed on to compute_offset()

Value

A data.frame suited for use in [PLN\(\)](#) and its variants with two special components: an abundance count matrix (in component "Abundance") and an offset vector/matrix (in component "Offset", only if offset is not set to "none")

Note

User supplied offsets should be either vectors/column-matrices or have the same number of column as the original count matrix and either (i) dimension names or (ii) the same dimensions as the count matrix. Samples are trimmed in exactly the same way to remove empty samples.

References

Chen, L., Reeve, J., Zhang, L., Huang, S., Wang, X. and Chen, J. (2018) GMPR: A robust normalization method for zero-inflated count data with application to microbiome sequencing data. PeerJ, 6, e4600 doi: [10.7717/peerj.4600](https://doi.org/10.7717/peerj.4600)

Paulson, J. N., Colin Stine, O., Bravo, H. C. and Pop, M. (2013) Differential abundance analysis for microbial marker-gene surveys. Nature Methods, 10, 1200-1202 doi: [10.1038/nmeth.2658](https://doi.org/10.1038/nmeth.2658)

Anders, S. and Huber, W. (2010) Differential expression analysis for sequence count data. *Genome Biology*, 11, R106 doi: [10.1186/gb20101110r106](https://doi.org/10.1186/gb20101110r106)

Kumar, M., Slud, E., Okrah, K. et al. (2018) Analysis and correction of compositional bias in sparse sequencing count data. *BMC Genomics* 19, 799 doi: [10.1186/s1286401851605](https://doi.org/10.1186/s1286401851605)

See Also

[compute_offset\(\)](#) for details on the different normalization schemes

Examples

```
data(trichoptera)
proper_data <- prepare_data(
  counts      = trichoptera$Abundance,
  covariates  = trichoptera$Covariate,
  offset      = "TSS"
)
proper_data$Abundance
proper_data$Offset
```

rPLN

PLN RNG

Description

Random generation for the PLN model with latent mean equal to mu, latent covariance matrix equal to Sigma and average depths (sum of counts in a sample) equal to depths

Usage

```
rPLN(
  n = 10,
  mu = rep(0, ncol(Sigma)),
  Sigma = diag(1, 5, 5),
  depths = rep(10000, n)
)
```

Arguments

n	the sample size
mu	vectors of means of the latent variable
Sigma	covariance matrix of the latent variable
depths	Numeric vector of target depths. The first is recycled if there are not n values

Details

The default value for mu and Sigma assume equal abundances and no correlation between the different species.

Value

a $n * p$ count matrix, with row-sums close to depths

Examples

```
## 10 samples of 5 species with equal abundances, no covariance and target depths of 10,000
rPLN()
## 2 samples of 10 highly correlated species with target depths 1,000 and 100,000
## very different abundances
mu <- rep(c(1, -1), each = 5)
Sigma <- matrix(0.8, 10, 10); diag(Sigma) <- 1
rPLN(n=2, mu = mu, Sigma = Sigma, depths = c(1e3, 1e5))
```

sigma.PLNfit

Extract variance-covariance of residuals 'Sigma'

Description

Extract the variance-covariance matrix of the residuals, usually noted

$$\Sigma$$

in PLN models. This captures the correlation between the species in the latent space.

Usage

```
## S3 method for class 'PLNfit'
sigma(object, ...)
```

Arguments

object an R6 object with class [PLNfit](#)
... additional parameters for S3 compatibility. Not used

Value

A semi definite positive matrix of size p , assuming there are p species in the model.

See Also

[coef.PLNfit\(\)](#), [standard_error.PLNfit\(\)](#) and [vcov.PLNfit\(\)](#) for other ways to access

$$\Sigma$$

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1 + offset(log(Offset)), data = trichoptera)
sigma(myPLN) ## Sigma
```

sigma.PLNmixturefit *Extract variance-covariance of residuals 'Sigma'*

Description

Extract the variance-covariance matrix of the residuals, usually noted

$$\Sigma$$

in PLN models. This captures the correlation between the species in the latent space. or PLNmixture, it is a weighted mean of the variance-covariance matrices of each component.

Usage

```
## S3 method for class 'PLNmixturefit'
sigma(object, ...)
```

Arguments

object an R6 object with class [PLNmixturefit](#)
 ... additional parameters for S3 compatibility. Not used

Value

A semi definite positive matrix of size p, assuming there are p species in the model.

See Also

[coef.PLNmixturefit\(\)](#) for other ways to access

$$\Sigma$$

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLNmixture(Abundance ~ 1 + offset(log(Offset)),
  data = trichoptera, control_main = list(iterates = 0)) %>% getBestModel()
sigma(myPLN) ## Sigma
```

stability_selection *Compute the stability path by stability selection*

Description

This function computes the StARS stability criteria over a path of penalties. If a path has already been computed, the function stops with a message unless `force = TRUE` has been specified.

Usage

```
stability_selection(
  Robject,
  subsamples = NULL,
  control = list(),
  force = FALSE
)
```

Arguments

<code>Robject</code>	an object with class <code>PLNnetworkfamily</code> , i.e. an output from <code>PLNnetwork()</code>
<code>subsamples</code>	a list of vectors describing the subsamples. The number of vectors (or list length) determines the number of subsamples used in the stability selection. Automatically set to 20 subsamples with size $10 \times \sqrt{n}$ if $n \geq 144$ and $0.8 \times n$ otherwise following Liu et al. (2010) recommendations.
<code>control</code>	a list controlling the main optimization process in each call to <code>PLNnetwork</code> . See <code>PLNnetwork()</code> for details.
<code>force</code>	force computation of the stability path, even if a previous one has been detected.

Value

the list of subsamples. The estimated probabilities of selection of the edges are stored in the fields `stability_path` of the initial `Robject` with class `PLNnetworkfamily`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
fits <- PLNnetwork(Abundance ~ 1, data = trichoptera)
## Not run:
n <- nrow(trichoptera)
subs <- replicate(10, sample.int(n, size = n/2), simplify = FALSE)
stability_selection(fits, subsamples = subs)

## End(Not run)
```

standard_error *Component-wise standard errors of Theta*

Description

Extracts univariate standard errors for the estimated coefficient of Theta. Standard errors are computed from the (approximate) Fisher information matrix. See [fisher.PLNfit\(\)](#) for more details on the approximations.

Usage

```
standard_error(object, type)

## S3 method for class 'PLNfit'
standard_error(object, type = c("wald", "louis"))
```

Arguments

object	an R6 object with class PLNfit
type	Either Wald (default) or Louis. Approximation scheme used to compute the Fisher information matrix

Value

A $p * d$ positive matrix (same size as Θ) with standard errors for the coefficients of Θ

Methods (by class)

- PLNfit: Component-wise standard errors of Theta in [PLNfit](#)

See Also

[vcov.PLNfit\(\)](#) for the complete Fisher information matrix

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1 + offset(log(Offset)), data = trichoptera)
standard_error(myPLN, "wald")
```

trichoptera

Trichoptera data set

Description

Data gathered between 1959 and 1960 during 49 insect trapping nights. For each trapping night, the abundance of 17 Trichoptera species is recorded as well as 6 meteorological variables which may influence the abundance of each species. Finally, the observations (that is to say, the trapping nights), have been classified into 12 groups corresponding to contiguous nights between summer 1959 and summer 1960.

Usage

trichoptera

Format

A list with 2 two data frames:

Abundance a 49 x 17 matrix of abundancies/counts (49 trapping nights and 17 trichoptera species)

Covariate a 49 x 7 data frame of covariates:

Temperature Evening Temperature in Celsius

Wind Wind in m/s

Pressure Pressure in mm Hg

Humidity relative to evening humidity in percent

Cloudiness proportion of sky coverage at 9pm

Precipitation Nighttime precipitation in mm

Group a factor of 12 levels for the definition of the consecutive night groups

In order to prepare the data for using formula in multivariate analysis (multiple outputs and inputs), use `prepare_data()`. We only kept a subset of the original meteorological covariates for illustration purposes.

Source

Data from P. Usseglio-Polatera.

References

Usseglio-Polatera, P. and Auda, Y. (1987) Influence des facteurs météorologiques sur les résultats de piégeage lumineux. *Annales de Limnologie*, 23, 65–79. (code des espèces p. 76) See a data description at <http://pbil.univ-lyon1.fr/R/pdf/pps034.pdf> (in French)

See Also

`prepare_data()`

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
```

vcov.PLNfit	<i>Calculate Variance-Covariance Matrix for a fitted PLN() model object</i>
-------------	---

Description

Returns the variance-covariance matrix of the main parameters of a fitted [PLN\(\)](#) model object. The main parameters of the model correspond to

$$\Theta$$

, as returned by [coef.PLNfit\(\)](#). The function can also be used to return the variance-covariance matrix of the residuals. The latter matrix can also be accessed via [sigma.PLNfit\(\)](#)

Usage

```
## S3 method for class 'PLNfit'
vcov(object, type = c("main", "covariance"), ...)
```

Arguments

object	an R6 object with class PLNfit	
type	type of parameter that should be extracted. Either "main" (default) for	Θ
	or "covariance" for	Σ
...	additional parameters for S3 compatibility. Not used	

Value

A matrix of variance/covariance extracted from the [PLNfit](#) model. If type="main" and Θ is a matrix of size $d * p$, the result is a block-diagonal matrix with p (number of species) blocks of size d (number of covariates). if type="main", it is a symmetric matrix of size p .

See Also

[sigma.PLNfit\(\)](#), [coef.PLNfit\(\)](#), [standard_error.PLNfit\(\)](#)

Examples

```
data(trichoptera)
trichoptera <- prepare_data(trichoptera$Abundance, trichoptera$Covariate)
myPLN <- PLN(Abundance ~ 1 + offset(log(Offset)), data = trichoptera)
vcov(myPLN) ## variance-covariance of Theta
vcov(myPLN, type = "covariance") ## Sigma
```

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