

# Package ‘pmd’

August 22, 2019

**Type** Package

**Title** Paired Mass Distance Analysis for GC/LC-MS Based Non-Targeted Analysis

**Version** 0.1.5

**Date** 2019-08-21

**Maintainer** Miao YU <yufreecas@gmail.com>

**Description** Paired mass distance (PMD) analysis proposed in Yu, Olkowicz and Pawliszyn (2018) <doi:10.1016/j.aca.2018.10.062> for gas/liquid chromatography–mass spectrometry (GC/LC-MS) based non-targeted analysis. PMD analysis including GlobalStd algorithm and structure/reaction directed analysis. GlobalStd algorithm could found independent peaks in m/z-retention time profiles based on retention time hierarchical cluster analysis and frequency analysis of paired mass distances within retention time groups. Structure directed analysis could be used to find potential relationship among those independent peaks in different retention time groups based on frequency of paired mass distances. A GUI for PMD analysis is also included as a 'shiny' application.

**URL** <https://yufree.github.io/pmd>

**BugReports** <https://github.com/yufree/pmd/issues>

**License** GPL-2

**Encoding** UTF-8

**LazyData** true

**Suggests** knitr, enviGCMS, igraph

**VignetteBuilder** knitr

**biocViews**

**Depends** R (>= 3.5.0)

**Imports** RColorBrewer, shiny, rmarkdown, rcdk, stats, utils

**RoxygenNote** 6.1.1

**NeedsCompilation** no

**Author** Miao YU [aut, cre] (<<https://orcid.org/0000-0002-2804-6014>>)

**Repository** CRAN

**Date/Publication** 2019-08-22 15:50:02 UTC

**R topics documented:**

getchain . . . . .	2
getcluster . . . . .	3
getcorcluster . . . . .	4
getpaired . . . . .	4
getpmd . . . . .	5
getrda . . . . .	6
getreact . . . . .	7
getsda . . . . .	8
getstd . . . . .	9
gettarget . . . . .	10
globalstd . . . . .	10
hmdb . . . . .	11
hmdbp . . . . .	12
keggrall . . . . .	12
omics . . . . .	13
pcasf . . . . .	13
plotpaired . . . . .	14
plotrtg . . . . .	15
plotsda . . . . .	15
plotstd . . . . .	16
plotstdrt . . . . .	17
plotstdsda . . . . .	17
runPMD . . . . .	18
sda . . . . .	18
spmein vivo . . . . .	19
<b>Index</b>	<b>20</b>

---

getchain	<i>Get reaction chain for specific mass to charge ratio</i>
----------	---

---

**Description**

Get reaction chain for specific mass to charge ratio

**Usage**

```
getchain(list, diff, mass, accuracy = 4, ...)
```

**Arguments**

list	a list with mzrt profile
diff	paired mass distance(s) of interests
mass	a specific mass for known compound or a vector of masses
accuracy	measured mass or mass to charge ratio in digits, default 4
...	other parameters for getpmd

**Value**

a list with mzrt profile and reaction chain dataframe

**Examples**

```
data(spmeinvivo)
# check metabolites of C18H39NO
pmd <- getchain(spmeinvivo,diff = c(2.02,14.02,15.99),mass = 286.3101)
```

---

getcluster

*Get Pseudo-Spectrum as peaks cluster based on pmd analysis.*

---

**Description**

Get Pseudo-Spectrum as peaks cluster based on pmd analysis.

**Usage**

```
getcluster(list, corcutoff = NULL, accuracy = 4)
```

**Arguments**

list	a list from getstd function
corcutoff	cutoff of the correlation coefficient, default NULL
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with Pseudo-Spectrum index

**See Also**

[getpaired](#),[getstd](#),[plotstd](#)

**Examples**

```
data(spmeinvivo)
re <- getpaired(spmeinvivo)
re <- getstd(re)
cluster <- getcluster(re)
```

---

getcorcluster	<i>Get Pseudo-Spectrum as peaks cluster based on correlation analysis.</i>
---------------	--

---

**Description**

Get Pseudo-Spectrum as peaks cluster based on correlation analysis.

**Usage**

```
getcorcluster(list, corcutoff = 0.9, rtcutoff = 10, accuracy = 4)
```

**Arguments**

list	a list with peaks intensity
corcutoff	cutoff of the correlation coefficient, default 0.9
rtcutoff	cutoff of the distances in cluster, default 10
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with Pseudo-Spectrum index

**Examples**

```
data(spmein vivo)  
cluster <- getcorcluster(spmein vivo)
```

---

getpaired	<i>Filter ions/peaks based on retention time hierarchical clustering, paired mass distances(PMD) and PMD frequency analysis.</i>
-----------	--

---

**Description**

Filter ions/peaks based on retention time hierarchical clustering, paired mass distances(PMD) and PMD frequency analysis.

**Usage**

```
getpaired(list, rtcutoff = 10, ng = NULL, digits = 2, accuracy = 4)
```

**Arguments**

list	a list with mzrt profile
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
ng	cutoff of global PMD's retention time group numbers, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with tentative isotope, multi-chargers, adducts, and neutral loss peaks' index, retention time clusters.

**See Also**

[getstd, getsda, plotpaired](#)

**Examples**

```
data(spmeinvivo)
pmd <- getpaired(spmeinvivo)
```

---

getpmd

*Get pmd for specific reaction*

---

**Description**

Get pmd for specific reaction

**Usage**

```
getpmd(list, pmd, rtcutoff = 10, corcutoff = NULL, digits = 2,
        accuracy = 4)
```

**Arguments**

list	a list with mzrt profile
pmd	a specific paired mass distances
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
corcutoff	cutoff of the correlation coefficient, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with paired peaks for specific pmd.

**See Also**

[getpaired](#), [getstd](#), [getsda](#), [getrda](#)

**Examples**

```
data(spmein vivo)
pmd <- getpmd(spmein vivo, pmd=15.99)
```

---

getrda

*Perform structure/reaction directed analysis for mass only.*

---

**Description**

Perform structure/reaction directed analysis for mass only.

**Usage**

```
getrda(mz, freqcutoff = 10, digits = 3, top = 20, formula = NULL)
```

**Arguments**

mz	numeric vector for independent mass or mass to charge ratio. Mass to charge ratio from GlobalStd algorithm is suggested. Isomers would be excluded automatically
freqcutoff	pmd frequency cutoff for structures or reactions, default 10
digits	mass or mass to charge ratio accuracy for pmd, default 3
top	top n pmd frequency cutoff when the freqcutoff is too small for large data set
formula	vector for formula when you don't have mass or mass to charge ratio data

**Value**

logical matrix with row as the same order of mz or formula and column as high frequency pmd group

**See Also**

[getsda](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
sda <- getrda(spmein vivo$mz[std$stdmassindex])
```

---

getreact	<i>Get quantitative paired peaks list for specific reaction/pmd</i>
----------	---

---

**Description**

Get quantitative paired peaks list for specific reaction/pmd

**Usage**

```
getreact(list, pmd, rtcutoff = 10, digits = 2, accuracy = 4,  
         ratiocv = 30, ...)
```

**Arguments**

list	a list with mzrt profile and data
pmd	a specific paired mass distances
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4
ratiocv	ratio cv cutoff for quantitative paired peaks, default 30
...	other parameters for getpmd

**Value**

list with quantitative paired peaks.

**See Also**

[getpaired](#), [getstd](#), [getsda](#), [getrda](#), [getpmd](#),

**Examples**

```
data(spmein vivo)  
pmd <- getreact(spmein vivo, pmd=15.99)
```

---

getsda *Perform structure/reaction directed analysis for peaks list.*

---

### Description

Perform structure/reaction directed analysis for peaks list.

### Usage

```
getsda(list, rtcutoff = 10, freqcutoff = 10, top = 50,  
       corcutoff = NULL, digits = 2, accuracy = 4)
```

### Arguments

list	a list with mzrt profile
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
freqcutoff	cutoff of frequency of PMDs between RT cluster for peaks, default 10
top	top n pmd frequency cutoff when the freqcutoff is too small for large data set, default 50
corcutoff	cutoff of the correlation coefficient, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

### Value

list with tentative isotope, adducts, and neutral loss peaks' index, retention time clusters.

### See Also

[getpaired](#), [getstd](#), [plotpaired](#)

### Examples

```
data(spmeinvivo)  
pmd <- getpaired(spmeinvivo)  
std <- getstd(pmd)  
sda <- getsda(std)
```



---

getstd	<i>Find the independent ions for each retention time hierarchical clustering based on PMD relationship within each retention time cluster and isotope and return the index of the std data for each retention time cluster.</i>
--------	---

---

### Description

Find the independent ions for each retention time hierarchical clustering based on PMD relationship within each retention time cluster and isotope and return the index of the std data for each retention time cluster.

### Usage

```
getstd(list, corcutoff = NULL, digits = 2, accuracy = 4)
```

### Arguments

list	a list from getpaired function
corcutoff	cutoff of the correlation coefficient, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

### Value

list with std mass index

### See Also

[getpaired](#), [getsda](#), [plotstd](#)

### Examples

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
```

`gettarget`*Get multiple injections index for selected retention time*

---

**Description**

Get multiple injections index for selected retention time

**Usage**

```
gettarget(rt, drt = 10, n = 6)
```

**Arguments**

`rt` retention time vector for peaks in seconds  
`drt` retention time drift for targeted analysis in seconds, default 10.  
`n` max ions numbers within retention time drift windows

**Value**

index for each injection

**Examples**

```
data(spmein vivo)  
pmd <- getpaired(spmein vivo)  
std <- getstd(pmd)  
index <- gettarget(std$rt[std$stdmassindex])  
table(index)
```

---

`globalstd`*GlobalStd algorithm with structure/reaction directed analysis*

---

**Description**

GlobalStd algorithm with structure/reaction directed analysis

**Usage**

```
globalstd(list, rtcutoff = 10, ng = 10, corcutoff = NULL,  
freqcutoff = 10, top = 50, digits = 2, accuracy = 4)
```

**Arguments**

<code>list</code>	a peaks list with mass to charge, retention time and intensity data
<code>rtcutoff</code>	cutoff of the distances in cluster, default 10
<code>ng</code>	cutoff of global PMD's retention time group numbers
<code>corcutoff</code>	cutoff of the correlation coefficient, default NULL
<code>freqcutoff</code>	cutoff of frequency of PMDs between RT cluster for independent peaks, default 10
<code>top</code>	top n pmd frequency cutoff when the freqcutoff is too small for large data set, default 50
<code>digits</code>	mass or mass to charge ratio accuracy for pmd, default 2
<code>accuracy</code>	measured mass or mass to charge ratio in digits, default 4

**Value**

list with GlobalStd algorithm processed data.

**See Also**

[getpaired](#), [getstd](#), [getsda](#), [plotstd](#), [plotstdsda](#), [plotstdrt](#)

**Examples**

```
data(spmein vivo)
re <- globalstd(spmein vivo)
```

---

hmdb	<i>A dataframe containing HMDB top 10000 unique accurate mass pmd and related reactions</i>
------	---

---

**Description**

A dataframe containing HMDB top 10000 unique accurate mass pmd and related reactions

**Usage**

```
data(hmdb)
```

**Format**

A dataframe with atoms numbers of C, H, O, N, P, S

**percentage** accuracy of atom numbers prediction

**pmd** pmd with two digits

---

hmdbp	<i>A list dataset containing HMDB unique accurate mass pmd analysis results</i>
-------	---

---

**Description**

A list dataset containing HMDB unique accurate mass pmd analysis results

**Usage**

```
data(hmdbp)
```

**Format**

A list with two vectors

**massp** all unique hmdb mass probability across all pmds

**pmdp** pmds probability across all unique hmdb mass

---

keggrall	<i>A dataframe containing reaction related accurate mass pmd and related reaction formula with KEGG ID</i>
----------	--

---

**Description**

A dataframe containing reaction related accurate mass pmd and related reaction formula with KEGG ID

**Usage**

```
data(keggrall)
```

**Format**

A dataframe with KEGG reaction, their realted pmd and atoms numbers of C, H, O, N, P, S

**ID** KEGG reaction ID

**pmd** pmd with three digits

---

omics	<i>A dataframe containing multiple reaction database ID and their related accurate mass pmd and related reactions</i>
-------	---

---

**Description**

A dataframe containing multiple reaction database ID and their related accurate mass pmd and related reactions

**Usage**

```
data(omics)
```

**Format**

A dataframe with reaction and their related pmd

**KEGG** KEGG reaction ID

**RHEA\_ID** RHEA\_ID

**DIRECTION** reaction direction

**MASTER\_ID** master reaction RHEA ID

**ec** ec reaction ID

**ecocyc** ecocyc reaction ID

**macie** macie reaction ID

**metacyc** metacyc reaction ID

**reactome** reactome reaction ID

**compounds** reaction related compounds

**pmd** pmd with two digits

**pmd2** pmd with three digits

---

pcasf	<i>Compare matrices using PCA similarity factor</i>
-------	---

---

**Description**

Compare matrices using PCA similarity factor

**Usage**

```
pcasf(x, y, dim = NULL)
```

**Arguments**

x	Matrix with sample in column and features in row
y	Matrix is compared to x.
dim	number of retained dimensions in the comparison. Defaults to all.

**Value**

Ratio of projected variance to total variance

**Author(s)**

Edgar Zanella Alvarenga

**References**

Singhal, A. and Seborg, D. E. (2005), Clustering multivariate time-series data. J. Chemometrics, 19: 427-438. doi: 10.1002/cem.945

**Examples**

```
c1 <- matrix(rnorm(16),nrow=4)
c2 <- matrix(rnorm(16),nrow=4)
pcasf(c1, c2)
```

---

plotpaired

*Plot the mass pairs and high frequency mass distances*

---

**Description**

Plot the mass pairs and high frequency mass distances

**Usage**

```
plotpaired(list, index = NULL, ...)
```

**Arguments**

list	a list from getpaired function
index	index for PMD value
...	other parameters for plot function

**See Also**

[getpaired](#), [globalstd](#)



**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdrt](#)

**Examples**

```
data(spmein vivo)
re <- getpmd(spmein vivo, pmd=78.9)
plotsda(re)
```

---

plotstd

*Plot the std mass from GlobalStd algorithm*

---

**Description**

Plot the std mass from GlobalStd algorithm

**Usage**

```
plotstd(list)
```

**Arguments**

`list` a list from `getstd` function

**See Also**

[getstd](#), [globalstd](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
plotstd(std)
```



---

plotstdrt	<i>Plot the std mass from GlobalStd algorithm in certain retention time groups</i>
-----------	--

---

**Description**

Plot the std mass from GlobalStd algorithm in certain retention time groups

**Usage**

```
plotstdrt(list, rtcluster, ...)
```

**Arguments**

list	a list from getstd function
rtcluster	retention time group index
...	other parameters for plot function

**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdsda](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
plotstdrt(std, rtcluster = 6)
```

---

plotstdsda	<i>Plot the std mass from GlobalStd algorithm in structure directed analysis(SDA) groups</i>
------------	--

---

**Description**

Plot the std mass from GlobalStd algorithm in structure directed analysis(SDA) groups

**Usage**

```
plotstdsda(list, index = NULL, ...)
```

**Arguments**

list	a list from getsda function
index	index for PMD value
...	other parameters for plot function

**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdrt](#)

**Examples**

```
data(spmeinvivo)
re <- globalstd(spmeinvivo)
plotstdsda(re)
```

---

runPMD

*Shiny application for PMD analysis*

---

**Description**

Shiny application for PMD analysis

**Usage**

```
runPMD()
```

---

sda

*A dataset containing common Paired mass distances of substructure, ions replacements, and reaction*

---

**Description**

A dataset containing common Paired mass distances of substructure, ions replacements, and reaction

**Usage**

```
data(sda)
```

**Format**

A data frame with 94 rows and 4 variables:

**PMD** Paired mass distances

**origin** potential sources

**Ref.** references

**mode** positive, negative or both mode to find corresponding PMDs

---

spmein vivo

*A peaks list dataset containing 9 samples from 3 fish with triplicates samples for each fish from LC-MS.*

---

**Description**

A peaks list dataset containing 9 samples from 3 fish with triplicates samples for each fish from LC-MS.

**Usage**

```
data(spmein vivo)
```

**Format**

A list with 4 variables from 1459 LC-MS peaks:

**mz** mass to charge ratios

**rt** retention time

**data** intensity matrix

**group** group information

# Index

## \*Topic **datasets**

- hmdb, 11
- hmdbp, 12
- keggrall, 12
- omics, 13
- sda, 18
- spmein vivo, 19

- getchain, 2
- getcluster, 3
- getcorcluster, 4
- getpaired, 3, 4, 6–9, 11, 14, 15
- getpmd, 5, 7
- getrda, 6, 6, 7
- getreact, 7
- getsda, 5–7, 8, 9, 11
- getstd, 3, 5–8, 9, 11, 16–18
- gettarget, 10
- globalstd, 10, 14–18

- hmdb, 11
- hmdbp, 12

- keggrall, 12

- omics, 13

- pcasf, 13
- plotpaired, 5, 8, 14, 16–18
- plotrtg, 15
- plotsda, 15
- plotstd, 3, 9, 11, 16, 16, 17, 18
- plotstdrt, 11, 16, 17, 18
- plotstdsda, 11, 17, 17

- runPMD, 18

- sda, 18
- spmein vivo, 19